

MINISTRY OF AGRICULTURE, FISHERIES AND FOOD
DIRECTORATE OF FISHERIES RESEARCH

FISHERIES RESEARCH TECHNICAL REPORT
NUMBER 97

**Collaborative UK Marine Mammal Project:
summary of data produced 1988-1992**

R. J. Law (compiler)

LOWESTOFT
1994

This report has been compiled by R. J. Law, BSc CChem MRSC who is a Grade 7 officer (Principal Scientific Officer) in Aquatic Environment Protection Division, Section 2, based at the Fisheries Laboratory, Burnham-on-Crouch.

Fish. Res. Tech. Rep., MAFF Direct. Fish. Res., Lowestoft, (97): 42pp.

© Crown copyright, 1994

Requests for reproduction of material contained in this report should be addressed to MAFF

CONTENTS

Page

1. Introduction	5
2. Guidelines for the postmortem and tissue sampling of cetaceans	5
2.1. Preface	5
2.2. Postmortem	6
2.2.1 <i>Introduction</i>	6
2.2.2 <i>Basic measurements</i>	6
2.2.3 <i>External examination</i>	7
2.2.4 <i>Examination of abdominal organs (except gastro-intestinal tract, pancreas and spleen)</i>	7
2.2.5 <i>Examination of organs of head, neck and thorax</i>	8
2.2.6 <i>Examination of the gastro-intestinal tract, pancreas and spleen</i>	9
2.3. Collection, storage and despatch of samples	9
2.3.1 <i>Labelling of samples</i>	9
2.3.2 <i>Blubber, liver and kidney samples for organochlorine and heavy metal analyses</i>	9
2.3.3 <i>Histological samples</i>	10
2.3.4 <i>Parasites</i>	10
2.3.5 <i>Teeth</i>	10
2.3.6 <i>Aqueous humour</i>	10
2.3.7 <i>Despatch of pathological specimens by post</i>	10
3. Analytical methods and quality control information	11
3.1. Determining the age of marine mammals	11
3.2. Trace metals in liver	11
3.2.1 <i>Summary of MAFF analytical methods and quality control data</i>	11
3.2.2 <i>Summary of SOAFD analytical methods and quality control</i>	12
3.3. Organochlorines in blubber	12
4. Results	13
4.1. Biological data	13
4.2. Trace metals in liver, kidney and stomach contents	13
4.3. Organochlorines in blubber and stomach contents	13
5. Summary and conclusion	13
6. References	29
7. Acknowledgements	30
8. Participating organisations	31
Appendix 1. Cetacean postmortem report form	32
Appendix 2. Summary of results derived from the analysis of reference materials certified for trace metals (MAFF)	41

1. INTRODUCTION

Robin J. Law (MAFF)

The seal epidemic of 1988 caused by phocine distemper virus led to the death of over 17 000 common (harbour) seals (*Phoca vitulina*) around the coasts of Europe (Dietz et al., 1989). Media reports at the time (and subsequent papers — see Brouwer *et al.*, 1989; Simmonds and Johnston, 1989) suggested that the severity of the outbreak was the result of North Sea pollution, and that high contaminant concentrations in the tissues of the seals weakened their resistance to disease. Subsequent analyses of blubber of seals from eastern and north-eastern England showed that concentrations were at the lower end of the range reported for seals from the North and Baltic Seas (Law *et al.*, 1989(a)). The discovery of high organochlorine concentrations in the blubber of a bottlenose dolphin (*Tursiops truncatus*) calf from Cardigan Bay which died in 1988 raised the level of concern (Morris *et al.*, 1989). Cardigan Bay is home to a resident population of bottlenose dolphins, and is generally regarded as a relatively clean area. Within the UK analyses of marine mammal tissues (mainly seals and porpoises) had been conducted in the 1970s and 1980s on samples collected in the 1970s, but at the time of the seal epidemic no data had been published for animals sampled later than 1978. There was therefore approximately a ten year gap in knowledge of contaminant concentrations for UK marine mammals, and no basis on which to assess possible effects. Accordingly a rapid, but somewhat piecemeal, programme of marine mammal analysis was undertaken within the laboratories of the Ministry of Agriculture, Fisheries and Food (MAFF) and the Scottish Office Agriculture and Fisheries Department (SOAFD). Although the data obtained made it clear that contaminants were not the cause of death, it was evident that a wider and more collaborative approach was required in order to establish the possible role of contaminants in promoting the susceptibility of marine mammals to disease. This programme, co-ordinated and partially funded by the Department of the Environment (DoE), started in 1990. It combines information on strandings with the provision of samples for pathological and chemical analyses. Primarily covering England and Wales, it involves staff from the Institute of Zoology (IOZ), the National Environmental Research Council (NERC) Sea Mammal Research Unit (SMRU), the Natural History Museum (NHM), the MAFF Fisheries Laboratory at Burnham-on-Crouch, and the SOAFD Marine Laboratory in Aberdeen. The addresses of these institutes are given in Section 8.

The main findings of this collaborative programme are being published elsewhere, in papers in the open scientific literature. The aim of this report is to make the contaminant dataset and relevant biological information available to other researchers, in a way that is usually impossible for a dataset of this size within the

space available for a journal article. The report includes details of the methodology employed, and in the case of the methods for the determination of trace metals and organochlorine compounds, of the associated analytical quality control (AQC) procedures. In the case of trace metal analyses undertaken by MAFF, we have also presented results obtained for standard materials during the period of study. Data are presented here for metals and/or organochlorines in tissues from 275 marine mammals; 205 cetaceans of nine species (comprising 137 harbour porpoises (*Phocoena phocoena*), 44 common dolphins (*Delphinus delphis*), 7 striped dolphins (*Stenella coeruleoalba*), 6 bottlenose dolphins, 4 white-sided dolphins (*Lagenorhynchus acutus*), 3 white-beaked dolphins (*Lagenorhynchus albirostris*), 2 long-finned pilot whales (*Globicephala melas*), 1 minke whale (*Balaenoptera acutorostrata*) and 1 Risso's dolphin (*Grampus griseus*); and 70 seals (40 grey seals *Halichoerus grypus* and 30 common seals).

2. GUIDELINES FOR THE POSTMORTEM AND TISSUE SAMPLING OF CETACEANS

Thijs Kuiken (IOZ) and John R. Baker (University of Liverpool (UL))

2.1 Preface

The postmortem examination of marine mammal carcasses is undertaken to determine the presence of diseases, if possible to establish the cause of death, and to collect tissue samples for a variety of laboratory analyses. Written guidelines help to ensure that all steps of the examination are carried out and that all the necessary samples are taken. They also serve to standardise the methods, and minimise the variation resulting from different people conducting postmortem examinations and collecting samples.

The diagnosis of a disease — loosely defined as any deviation from the normal structure or function of any part of the body — is often based on the combined results of gross examination of organs, microscopical examination of diseased tissues, and other laboratory analyses, such as bacteriological examination. Once these results have been integrated and an overview has been obtained of the disease processes that are present in a marine mammal carcass, one can attempt to establish the cause of death. In some cases this is straightforward, for example when an animal has bled to death from a stomach ulcer and the intestines are filled with blood. In other cases it is not possible to determine which of the diseases found, if any, caused the animal's demise.

Postmortem examinations on cetaceans and seals were carried out according to protocols written by Kuiken and Baker (1991(a), 1991(b)). Several veterinary surgeons, based in different parts of the UK, undertook examinations for this programme and we acknowledge these in Section 7. To conform with a European protocol published by the European Cetacean Society (Kuiken and Garcia Hartmann, 1993), the guidelines for cetaceans have been revised and updated (Kuiken and Baker, 1993). These revised guidelines are reproduced in full below (Sections 2.2 and 2.3).

2.2 Postmortem

2.2.1 Introduction

These guidelines are meant primarily as an aid to veterinary surgeons carrying out postmortem examinations on stranded cetaceans in the UK, as a part of the DoE-funded marine mammal projects in England, Wales, and Scotland. They are based partly on guidelines written by Dr John Baker, University of Liverpool, and partly on the protocol produced at the European Cetacean Society workshop on cetacean pathology, held in Leiden, The Netherlands, in September 1991 (Kuiken and Garcia Hartmann, 1993).

All structures must be examined visually and by palpation, making incisions into the organs. A full postmortem record must be kept, preferably on the standard 'cetacean postmortem report' form (see Appendix 1).

Lesions in any organs should be described, photographed and sampled. The description should include the size, location, colour, texture, shape, and the nature of the transition from normal to abnormal tissue. Photographs should include a ruler or similar object to indicate the size of the lesion. According to the suspected aetiology of the lesion, samples should be collected for bacteriological examination (especially if the lesion is of a purulent nature), for virological examination, and for parasitological examination. In all

cases, a sample of the lesion should be preserved for histopathological examination.

Any parasites found, regardless if they are associated with pathological lesions or not, should be preserved in 70% ethanol for identification. An attempt should be made to estimate the total number of parasites. Some predilection sites for parasites are indicated in the text.

If the state of decomposition of the carcass is advanced (condition code 4 or 5, see below), only the basic measurements, organ weights (when possible), and a limited number of samples (epidermis, skull, teeth, food remains, gonads) should be taken.

The postmortem examination need not take place in the order described below. However, samples for bacteriological and virological examination need to be taken as early as possible. Also, examination of the gastrointestinal tract should be left until last to prevent cross-contamination with enteric micro-organisms.

2.2.2 Basic measurements

Photographs should be taken of the lateral views of the whole body, from both sides. Particularly in bottle-nosed dolphins, photographs should be taken of the dorsal fin, also from both sides. In baleen whales, the ventral side of the tail flukes should be photographed.

Estimate the body condition, that is the state of decomposition of the carcass, using the categories of the condition code¹.

Weigh the carcass. If this is not possible, the body weight can be estimated from the heart weight².

Measure the body length by placing the carcass on its belly, holding a measuring tape or ruler next to the carcass in a straight line parallel to the longitudinal body axis and measuring the distance between the notch in the tail flukes and the tip of the upper jaw. Measure the body girth at the level of the anterior insertion of the dorsal fin.

Notes:

¹ The body condition, or state of decomposition of a carcass, can be described using the following condition code:

- 1) **live** (becomes code 2 at death)
- 2a) *extremely fresh* (as if just died, no bloating, meat is considered by most to be edible)
- 2b) *slight decomposition* (slight bloating, blood imbibition visible)
- 3) **moderate decomposition** (bloating, skin peeling, penis may be extended in males, organs still intact, excluding postmortem damage)
- 4) **advanced decomposition** (major bloating, skin peeling, penis extended in males, organs beyond recognition, bones exposed due to decomposition)
- 5) **indeterminate** (mummified carcass or skeletal remains, no organs present)

² The body weight can be estimated from the heart weight using the formula
 $\log W = (\log H + 2.2) / 0.984$, with H = heart weight and W = body weight, both in kg.

2.2.3 External examination

Indicate the nutritional state of the carcase, using one of the following 3 categories:

- good: the aspect of the upper flanks on either side of the dorsal fin is rounded;
- moderate: the aspect of the upper flanks on either side of the dorsal fin is sloping;
- poor: the aspect of the upper flanks on either side of the dorsal fin is hollow (in these animals, one can make out the transverse processes of the lumbar vertebrae, and there is an indentation dorsally just behind the head).

Examine the body orifices (mouth, eyes, ear openings, blow-hole, anus, genital slit and mammary slits) for lesions and any discharge.

Examine the animal for external lesions and sample these accordingly. Examine the skin carefully for any ectoparasites. These are most likely to be found in or near the body orifices and next to the fins and flukes. Take a 4 cm² piece of epidermis down to the blubber for DNA-studies, and freeze.

Massage the skin in the area cranial to the mammary slits in a caudal direction to express any fluid present in the mammary glands. If fluid can be pressed out, collect a sample for organochlorine analysis in a hexane-washed glass container and freeze. If the lid is made of plastic, separate the sample from this with aluminium foil. Note the volume, colour, and consistency of the fluid.

Cut a transverse strip of blubber about 2 cm wide from the anterior insertion of the dorsal fin, from the mid-dorsal to the mid-ventral region. Make sure to cut at right-angles to the surface of the skin. Measure the thickness of the blubber strip with a ruler 2 cm lateral to the dorsal mid-line, mid-laterally, and 2 cm lateral to the ventral mid-line. (Using this method, the tension of the blubber tissue is relieved before measuring.) In the harbour porpoise, take about 20 g of the lateral part of the blubber strip (preferably with some muscle tissue and the skin attached) and freeze. This sample is for lipid analysis.

Cut a strip of blubber a few cm wide and a few cm long at the level of the caudal insertion of the dorsal fin. Make sure to cut at right-angles to the surface of the skin. Measure the thickness of the blubber strip 2 cm lateral to the dorsal mid-line. From this blubber strip, take 2 x 20 g cross-sectional samples of blubber for organochlorine analysis. It is important to take samples of the whole layer, from the skin to the muscle. Wrap them in hexane-washed aluminium foil, place them in 25 ml Universal tubes, and freeze. Alternatively, they can be placed in Sovirel glass tubes.

Take 2 x 20 g muscle samples for toxicological analysis, at the same location as and directly below the blubber sample, at the level of the caudal insertion of the dorsal fin. Wrap them in hexane-washed aluminium foil, place them in 25 ml Universal tubes, and freeze. Alternatively, they can be placed in Sovirel glass tubes.

With the animal on its right side make a mid-line ventral incision from the symphysis of the mandible to a short distance posterior of the anus, circumventing the genital slit and anus. From the posterior end of the ventral incision make a second one almost to the dorsal mid-line. Reflect the skin and blubber off the uppermost side. Any parasites in the blubber should be noted and collected. They may occur as white cysts less than 1 cm in diameter, often in the ano-genital region or the dorsal aspect of the chest wall.

In females, examine the mammary gland for pathological changes and parasites. Collect a cross-sectional slice of about 1 cm thick from halfway along the length of the left mammary gland for histopathological examination, and place in 10% formalin.

Examine the subcutaneous tissue for the presence of bruises and parasites.

Remove the left scapula for (future) radionuclide analysis and freeze.

2.2.4 Examination of abdominal organs (except gastro-intestinal tract, pancreas, and spleen)

Remove the left abdominal wall, freeing the testis or ovary and uterus. Any parasites in the abdominal wall (for instance cysts under the peritoneum) should be collected. Remove the left thoracic wall, for example with bone shears. Remove the fifth left rib and freeze a 15 cm section of it for (future) lead analysis.

Before handling the internal organs, take a 1 cm³ sample of lung tissue from the cranio-ventral part of the left lung and a 1 cm³ sample of kidney tissue from the left kidney for virological examination. Also take a sample of lung tissue from the cranio-ventral part of the left lung, a sample of kidney tissue from the left kidney, a sample of liver tissue from the left lobe of the liver, and a sample of heart blood from the right ventricle, for bacteriological examination.

Sever the intestine close to the anus and the oesophagus close to the diaphragm. Working forward along the dorsal aspect of the abdominal cavity, remove the stomach, intestines, pancreas, spleen and mesenteric lymph node, attached to each other, from the carcase. Leave the examination of the G.I. tract to the end of the postmortem examination to prevent cross-contamination of other tissues with enteric micro-organisms.

Open and examine the bladder *in situ*, noting the contents, if any.

In females remove the entire reproductive tract, open the vagina and uterus, note any corpora lutea, corpora albicantia or follicles on each ovary and then place the ovaries in 10% formalin for reproduction studies.

If a foetus is present of sufficient size to examine the individual organs, a postmortem examination and tissue sampling of the foetus can take place in the same way as for cetaceans after birth. If it is too small for a full postmortem examination, the whole foetus and its placenta can be wrapped in hexane-washed aluminium foil and stored frozen for organochlorine analysis.

In males remove the testes and weigh them separately after removing the epididymis. After incision and examination, place the testes in 10% formalin for reproductive studies. If they are heavier than about 50 g each, place a cross-sectional slice about 1 cm thick from mid-way along the length in 10% formalin. Examine the penis and preputium.

Remove and examine the adrenal glands, and place them separately in 10% formalin for morphometrics studies.

Remove the kidneys from the body cavity. Incise both kidneys longitudinally, and if possible, strip the capsule. Then, take 2 x 20 g samples for toxicological analysis from halfway the length of the left kidney. These samples should be cross-sectional and include both medullary and cortical tissue. Wrap them in hexane-washed aluminium foil, place them in 25 ml Universal tubes, and freeze. Alternatively, they can be placed in Sovirel glass tubes. Preserve 1 cm³ from a kidney in 10% formalin for histopathological examination.

Remove the liver, examine both surfaces and make multiple incisions into the substance. Examine the bile ducts for parasites. Then, take 2 x 20 g for trace metal analysis. These samples should include approximately equal amounts of tissue from the edge of the left lobe, the edge of the right lobe, and the hilus of the liver. Wrap them in hexane-washed aluminium foil, place them in 25 ml Universal tubes, and freeze. Alternatively, they can be placed in Sovirel glass tubes. Place 1 cm³ of liver tissue in 10% formalin for histopathological examination.

2.2.5 Examination of organs of head, neck and thorax

Carefully remove the superficial muscles overlying the trachea and larynx, so exposing the thyroid gland. Examine this tissue.

Incise along the internal aspects of both mandibles and free the tongue. Once the tongue is free reflect it backwards and cut the hyoid bones close to the skull.

Free the larynx from the sphincter muscle holding it in place and pulling the tongue backwards incise along the neck to free the trachea and oesophagus. Then, incising dorsally and ventrally in the thoracic cavity, free the heart and lungs. Note any attachments of the lungs to the thoracic walls. This procedure should give you the tongue, larynx, trachea, oesophagus, thymus, heart and lungs all still fastened together.

Examine the surface of the tongue.

Open the oesophagus longitudinally and check for lesions or parasites.

Open the larynx, trachea and major bronchi longitudinally. Make multiple incisions into the substance of both lungs. Any parasites should be collected. Two pieces of lung (about 1 cm³) from the hilus and periphery of the left lung, and the same from the right lung, should be collected in 10% formalin for histopathological examination. The samples should include part of the major bronchial tree. Repeat this procedure, placing the second set of samples in a separate container with 10% formalin for morbillivirus detection using an immunoperoxidase test. Open all major branches of the pulmonary veins and examine for the presence of parasites. Examine the bronchial and so-called 'pulmonary associated' lymph nodes. The latter can be found about halfway along the ventral edges of each lung. Cut a 1 cm thick cross-sectional slice from the middle of the left pulmonary associated lymph node, and place it in 10% formalin for histopathological examination.

Examine the thymus, if present and place 1 cm³ in 10% formalin for histopathological examination. Freeze another 1 cm³ of the thymus for immunocytological examination, if the carcass is very fresh (condition code 2).

Collect any blood present in the heart lumen, to obtain serum for serological examination. The serum, acquired by centrifugation, should be stored frozen. Even if it is haemolytic, it is still of value.

Separate the heart from the lungs by cutting through the major blood vessels where they enter the heart. Open the left and right ventricles and atria for examination and to take out any blood clots present. Any parasites should be collected. Weigh the heart. Cut a 1 cm thick slice of heart tissue, to include a piece of the wall of the left ventricle and of the atrioventricular septum, and place it in 10% formalin for histopathological examination.

With a saw or bone shears break open the tympanic bullae (which in cetaceans are not part of the skull but lie free just behind the mandibles). With forceps examine the internal cavity to recover any nematodes present.

If possible, remove at least 4 teeth from the middle of the lower jaw for aging, and store frozen. (In baleen whales, cut off 2 baleen plates as near as possible to their basis and store frozen.)

In freshly dead carcasses (condition code 2), open the skull, and examine the brain.

The skull can be opened by making a vertical cut parallel and about 2 cm posterior to the transverse dorsal ridge which is clearly visible and palpable on top of the skull. The second cut should be made in the horizontal plane, through the occipital condyles, making sure to leave the posterior portion of the condyles on the skull, so that the condylo-basal length can still be measured. Both cuts should be extended until they meet each other. The separated piece of skull can then be pried loose using a chisel or flat-bladed screwdriver, and the brain can be removed.

Take a 1 cm³ sample of brain for virological examination. Place the rest of the brain in 10% formalin for at least a week. To allow faster fixation, a longitudinal incision can be made in the cerebrum to expose the lateral ventricles. When it is fixed, make multiple slices into the tissue to look for pathological lesions, including the presence of parasites. Take 1 cm³ samples of the cortex, midbrain, cerebellum, and medulla, for histopathological examination.

In more decomposed carcasses, leave the skull intact. Both opened and completely intact skulls should be stored frozen for morphometrics studies.

2.2.6 Examination of the gastro-intestinal tract, pancreas and spleen

Examine the spleen and put a piece (about 1 cm³), including a section of capsule, in 10% formalin for histopathological examination. One often finds smaller accessory spleens near to the main spleen.

Examine the pancreas. Look for parasites, particularly in the pancreatic ducts. Place a 1 cm³ piece of pancreas tissue in 10% formalin for histopathological examination.

Examine the mesenteric lymph node and put a 1 cm thick cross-sectional slice from halfway along its length in 10% formalin for histopathological examination.

Open the cardiac section of the stomach. Weigh any fish bones, otoliths and other food remains and preserve in 70% ethanol or freeze for prey studies. Any parasites should be collected. Describe any lesions, including the distribution and size of any ulcers.

Open the fundic and pyloric sections of the stomach. Any food material and parasites should be preserved as for the cardiac section. Any nodules in the walls of the fundic and pyloric sections should be noted and, if they are found, attempts should be made to express the contents. Any parasites found in the contents should be collected.

Free the intestine from the mesentery and open the entire length of the organ collecting any contents in a

bucket. The contents should be diluted with water and sieved through a 500 µm sieve to collect otoliths and other recognisable food remains. These should be stored in 70% ethanol (or alternatively stored frozen). Any parasites should be collected.

2.3 Collection, storage and despatch of samples

2.3.1 Labelling of samples

As the samples from each animal are to be examined by a number of people, it is very important that the samples are properly labelled. This means that:

- (i) each sample should have a label which is securely fastened to it.
- (ii) indelible ink or pencil should be used to write on the labels.
- (iii) the label should include the Natural History Museum reference number of the carcass (if not available your own reference number) and the name of the sample (e.g. SW1990/56 aqueous humour).

2.3.2 Blubber, liver and kidney samples for organochlorine and heavy metal analyses

Collection

It is important to take the blubber sample of the whole layer, from the skin to the muscle.

Blubber samples should only come into contact with stainless steel, aluminium, glass or teflon. The glass Sovirel tubes provided by the MAFF laboratory in Burnham-on-Crouch have tops with a teflon lining on the inside. Make sure this lining is in place before use. The glass has been washed in hexane to remove any remains of plastics.

If Sovirel tubes are not available, the blubber, liver and kidney samples can also be wrapped in aluminium foil.

Storage and package

If samples are sent by post directly after collection, they do not need to be cooled. Otherwise, they should be frozen at -20°C and not thawed during storage, otherwise the tissue will break down. To send by post, the tubes should be suitably wrapped to prevent breakage and placed in an insulating container, such as polystyrene. They should be sent by a fast postal service (e.g. Redstar, Datalink), at the beginning of the week.

2.3.3 Histological Samples

Fixation

10% neutral-buffered formalin should be used as a fixative. This can be made as follows: dissolve 3.31 g $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$ and 33.77 g $\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$ in 1 l distilled water. To 1 part of concentrated formalin (= 40% formaldehyde solution) add 9 parts of this buffer.

As a rule, formalin will only penetrate about 1 cm in any direction, so inject or cut samples larger than 2 cm in diameter to expose more tissue to the formalin. Also remember to try to have 10 times the volume of 10% formalin to tissue when fixing.

Storage and package

Once fixation has taken place, the samples can be stored in a smaller volume of 10% formalin. Samples should be posted in well-sealed plastic vials.

2.3.4 Parasites

Fixation

Live specimens should be immersed for about 1 minute in Berland's fluid (19 parts pure glacial acetic acid + 1 part concentrated formalin (= 40% formaldehyde solution)) and then stored in 70-80% ethanol. Dead specimens can be put in 70-80% ethanol straightaway.

Storage and package

Specimens should be stored in 70% ethanol, and sent by post in strong, well-sealed plastic vials.

2.3.5 Teeth

Collection

Most toothed cetaceans can be aged by examining the growth layers of the teeth, so it is always worthwhile to collect teeth for age determination. At least 2, and preferably 4, teeth should be collected from each individual³. From the common porpoise, teeth can easily be extracted by inserting a sharp knife or scalpel, firstly on one side of the gum in between the teeth and the connective tissue, and then on the other side. With a little pulling an entire string of teeth can be removed. In species such as the bottlenose dolphin, the teeth can be loosened by prying in between the space between the tooth and the socket with a sharp pointed instrument. If this is not possible, a piece of mandible containing at least 4 teeth can be sawed off. It doesn't matter which teeth are taken, as long as they are damaged or worn as little as possible. It is important to extract the tooth in its entirety, i.e., including all the root.

Storage and package

Loose teeth and jaw sections can be stored frozen. Alternatively, they can be fixed in 70% alcohol or 10% neutral-buffered formalin. They should not be stored dry as this may lead to cracking of the teeth. The samples can be sent by post in well-sealed plastic vials, or wrapped in wet tissue-paper and then placed in a plastic bag.

2.3.6 Aqueous humour

Collection

Put pressure on one corner of the eye with a finger. Using a syringe and thin hypodermic needle (i.e. 23 gauge), puncture the eye bulb at the other corner of the eye and extract the aqueous humour. (In porpoises, approximately 0.75 ml of fluid can be collected from each eye in this manner.)

Storage and package

Store frozen. Samples should be posted in well-sealed plastic vials, placed in a suitable isolating container.

2.3.7 Despatch of pathological specimens by post

In order to assure rapid and safe delivery of samples and to comply with Post Office rules, the following points should be adhered to:

Only first-class letter post or data post may be used. Parcel post must not be used.

Every specimen must be enclosed in a primary container which is securely sealed.

The primary container must be wrapped in sufficient absorbent material to absorb all possible leakage in the event of damage.

The container and absorbent material must be sealed in a leakproof plastic bag. In the case of frozen specimens, this package should be placed in a suitable isolating container, such as polystyrene.

This must then be placed in a strong box or case in such a way that it cannot move about. If necessary the box/case should be placed in a stout envelope or padded bag.

The completed package must be clearly marked 'Pathological Specimen - Fragile. With Care' and show the name and address of the sender to be contacted in case of leakage.

*Note:*³ With beaked whales it may only be possible to collect one tooth

A statement detailing the nature of the contents must accompany all specimens. The statement must be so concealed as to easily allow the consignee to identify the nature of the specimen before breaking into the inner protective packaging.

3. ANALYTICAL METHODS AND QUALITY CONTROL INFORMATION

3.1 Determining the age of marine mammals

Christina Lockyer (SMRU)

Whenever possible at least 4 teeth were removed from the middle of the lower jaw for aging purposes, and stored frozen (see Section 2.2.5 above). The teeth were decalcified, sectioned, stained and mounted on microscope slides. The sectioned and stained teeth were then examined under a binocular microscope and the growth layers in dentine and / or cementum counted. A fuller outline of the procedure can be found in Lockyer (in press).

3.2 Trace metals in liver

Bryn R. Jones (MAFF) and John Pirie (SOAFD)

The majority of the trace metal analyses (all but twenty samples) were conducted by MAFF, the remainder by SOAFD.

3.2.1 Summary of MAFF analytical methods and quality control data

Samples of marine mammal liver are weighed into a microwave digestion vessel, (~2.5 g wet), and the sample is dried at 102°C for 12 hours. After cooling the sample is reweighed and the weight of dry material determined. 4 cm³ of concentrated nitric acid and 4 cm³ distilled water are added to the vessel, then the vessel is closed and the sample digested in an analytical microwave oven (Model MDS-2000, CEM Corporation, Matthews, North Carolina, USA) for 25 minutes, during which the temperature and pressure in the digestion vessels reach 120°C and 120 psi respectively. After cooling, the samples are quantitatively transferred to polypots and made up to 25 cm³ volume using distilled water.

Most of the batches of sample were analysed by the following procedures:

Prior to analysis hydroxylammonium chloride was added to the digest in order to remove excess nitric acid.

For all instrumental analysis, suitable external calibrations are made and the digests diluted to read within the calibration range. Samples were analysed for nickel, copper, cadmium and zinc by atomic absorption spectrophotometry (AAS) with an air/acetylene flame (Varian SpectrAA-400P flame instrument; Varian Ltd., Walton-on-Thames, Surrey). Chromium was analysed using the same instrument with a nitrous oxide/acetylene flame. Lead was analysed by electrothermal AAS using a Pye-Unicam SP9 spectrophotometer with graphite furnace employing pyrolytic graphite coated tubes. Mercury was analysed by atomic fluorescence following reduction by tin II chloride using a dedicated system (Merlin; P. S. Analytical Ltd., Sevenoaks, Kent).

The determination of arsenic requires the use of a separate digestion procedure. Samples for arsenic were digested using the procedure for arsenic in fish tissue given by Harper *et al.* (1989), the final analysis being carried out using the Varian instrument described above in conjunction with a Varian VG 76 continuous flow hydride generator.

The determination of selenium also requires a separate digestion procedure. 1.5 g of liver was weighed into an open beaker. Concentrated nitric and hydrochloric acids were added (10 cm³ each) and the samples were heated on hot plates until a clear digest was obtained (4 - 8 hours). Samples were quantitatively made up to volume with 10% hydrochloric acid. The instrumentation used for the final analysis was as for arsenic above.

Following the purchase of new instrumentation and appropriate method development the final batches of samples were analysed for metals other than mercury by means of inductively-coupled plasma/mass spectrometry (ICP/MS) (VG PlasmaQuad PQ2+; FI Elemental, Winsford, Cheshire). In this technique the standard nitric acid digest is suitable for all elements, and external calibration is used in conjunction with internal standards to compensate for instrumental drift and matrix interferences. Arsenic and selenium also require corrections to be made for chloride interferences.

No significant biases have been observed between data generated using AAS and ICP/MS techniques. Prior to the commencement of analyses of marine mammal tissue we participated successfully in an international intercomparison exercise in which narwhal liver was analysed for trace metals (Wagemann and Armstrong, 1988). All analyses reported here were conducted under a Quality Assurance (QA) programme under which standard materials and procedural blanks were included in each batch of samples analysed. The results obtained from analyses of certified reference materials analysed in 1991 and 1992 are summarised in Appendix 2. Full details of the current methodology and the associated AQC procedures will be given in Jones and Laslett (in prep.).

A number of kidney samples from a range of marine mammals, and a number of stomach content samples from common dolphins were also analysed in conjunction with studies of a mass mortality in the south-west of England that occurred early in 1992. These samples were treated in essentially the same way as the liver samples.

3.2.2 Summary of SOAFD analytical methods and quality control

Tissue samples (0.5 - 1.0 g wet weight) were digested in silica flasks with 10 cm³ of concentrated nitric acid (Aristar grade). The acid volume was reduced to 5 cm³, and the digest allowed to cool before being quantitatively transferred to a 25 cm³ calibrated plastic vial and made up to volume with distilled water. Analyses for copper and zinc were by AAS with an air/acetylene flame (model 5000; Perkin Elmer, Beaconsfield, Bucks.). Cadmium, chromium, lead and selenium were analysed by electrothermal AAS with heated graphite atomisation (Perkin Elmer Zeeman 3030). Pyrolytic graphite coated tubes with a L'Vov platform, in conjunction with a matrix modifier and thermal programme, were employed to effect atomisation. Mercury was determined by cold vapour AAS (Data Acquisition Mercury Flow Meter) following reduction with tin II chloride. All instruments were calibrated using acid matrix matched standards, with a minimum of five standards per line. The methods were validated by analysis of appropriate reference materials, and participation in intercalibration exercises. Quality control was maintained by analysis of the certified reference material DOLT-1 — all the samples reported here were run in a single batch.

3.3 Organochlorines in Blubber Colin R. Allchin (MAFF)

Blubber samples were taken according to the protocol in Section 2.2.3. Cross-sectional samples of blubber adjacent to the dorsal fin were collected with a stainless steel knife, placed in a hexane-washed glass container (generally a Sovirel tube) with a PTFE- or aluminium-lined cap, and stored at -20°C until analysis was carried out. The concentrations of the following contaminants were determined according to the procedures outlined by Allchin *et al.* (1989): hexachlorobenzene (HCB), α -HCH (hexachlorocyclohexane), γ -HCH (lindane), *p,p'*-DDE, *p,p'*-TDE, *p,p'*-DDT, dieldrin, and 25 individual chlorobiphenyl congeners numbered according to Ballschmiter and Zell (1980), and listed in Table 1. To obtain the desired chromatographic separations analyses were routinely conducted on two columns of differing polarity and selectivity: 50 m x 0.2 mm ID HP-5 (5% phenyl methylsilicone; film thickness 0.33 μ m) and 50 m x 0.25 mm ID CP-Sil 19-CB (film thickness 0.25 μ m). In both cases hydrogen was used as the carrier gas, and detection was by means of an electron-

capture detector. The columns were mounted in HP5890 gas chromatographs, and injections were made automatically by means of HP7673a autosamplers. A range of alkyl substituted dichlorobenzylethers (C₆, C₇, C₁₄ and C₁₆) were used as internal standards, and were added to the samples prior to GC analysis of the extracts. Some confirmatory analyses were also conducted by coupled gas chromatography/mass spectrometry using a Finnigan-MAT ITS-40 instrument.

Table 1. List of chlorobiphenyl congeners analysed in blubber samples

Congener number	Structure
CB18	2,2',5'-trichlorobiphenyl
CB28 *	2,4,4'-trichlorobiphenyl
CB31	2,4',5'-trichlorobiphenyl
CB44	2,2',3,5'-tetrachlorobiphenyl
CB47	2,2',4,4'-tetrachlorobiphenyl
CB49	2,2',4,5'-tetrachlorobiphenyl
CB52 *	2,2',5,5'-tetrachlorobiphenyl
CB66	2,3',4,4'-tetrachlorobiphenyl
CB101 *	2,2',4,5,5'-pentachlorobiphenyl
CB105	2,3,3',4,4'-pentachlorobiphenyl
CB110	2,3,3',4',6-pentachlorobiphenyl
CB118 *	2,3',4,4',5-pentachlorobiphenyl
CB128	2,2',3,3',4,4'-hexachlorobiphenyl
CB138 *	2,2',3,4,4',5'-hexachlorobiphenyl
CB141	2,2',3,4,5,5'-hexachlorobiphenyl
CB149	2,2',3,4',5',6-hexachlorobiphenyl
CB151	2,2',3,5,5',6-hexachlorobiphenyl
CB153 *	2,2',4,4',5,5'-hexachlorobiphenyl
CB156	2,3,3',4,4',5-hexachlorobiphenyl
CB158	2,3,3',4,4',6-hexachlorobiphenyl
CB170	2,2',3,3',4,4',5-heptachlorobiphenyl
CB180 *	2,2',3,4,4',5,5'-heptachlorobiphenyl
CB183	2,2',3,4,4',5',6-heptachlorobiphenyl
CB187	2,2',3,4',5,5',6-heptachlorobiphenyl
CB194	2,2',3,3',4,4',5,5'-octachlorobiphenyl

The suite of CBs comprises 3 trichlorobiphenyls, 5 tetrachlorobiphenyls, 4 pentachlorobiphenyls, 8 hexachlorobiphenyls, 4 heptachlorobiphenyls and 1 octachlorobiphenyl. The seven congeners comprising the ICES primary list are indicated by asterisks

Sample preparation and fractionation were conducted at SMRU, with the chromatographic analysis and interpretation being undertaken at MAFF. Procedural blanks were run with all sample batches, and in addition all the solvents and reagents (alumina, silica, anhydrous sodium sulphate) were checked for contamination prior to use. A reference fish-oil (SOAFD LRM-1) was also analysed with each batch to provide AQC information on the performance of the methods, and two certified reference materials (CRM 349 and 350: cod liver and mackerel oils respectively) from the EC Community Bureau of Reference were analysed periodically.

A number of stomach content samples from common dolphins were also analysed in conjunction with studies of a mass mortality in the south-west of England. These were treated in essentially the same way as the blubber samples.

4. RESULTS

4.1 Biological data

Thijs Kuiken, Peter M. Bennett (IOZ), Christina Lockyer (SMRU), Sarah Phillips and Chris Spurrier (NHM)

The basic information for each of the animals for which data are reported is given in Tables 2 and 3. Age, where given, is in years. The body length is given in cm, and was defined as the straight length measurement from the tip of the upper jaw to either the tip of the tail (for seals), or to the tail notch (for cetaceans). Table 2 includes information for all animals other than harbour porpoises, and Table 3 that for porpoises alone. The main identifier for animals used is the NHM number (Natural History Museum stranding incident reference number) whenever possible, as this provides a link to other information. Where no NHM number was assigned another unique identifier has been created. The basic information comprises species, sex, age, length, and date and location where found, where each of these pieces of information is known. Dashes indicate where information is unavailable. Table 3 also indicates the probable cause of death as established by postmortem investigations of harbour porpoise carcasses.

4.2 Trace metals in liver, kidney and stomach contents

Bryn R. Jones (MAFF) and John Pirie (SOAFD)

Table 4 lists the trace metal concentrations observed in samples of liver tissue, Table 5 those for kidney tissue, and Table 6 to analyses of stomach contents from a particular group of common dolphins. In all cases the concentrations are given in units of mg kg⁻¹ wet weight, although the percentage dry matter in the samples is also given to facilitate conversion to dry weight if desired. In all tables the reference numbers are given for each animal in order to provide a link to Tables 2 and 3. In Table 4 two dolphins are listed with the same reference number (IC1991/1); the animal was a pregnant female and the suffix A refers to the mother, suffix B to her foetus (as in Table 2).

4.3 Organochlorines in blubber and stomach contents

Colin R. Allchin (MAFF) and Mike Walton (SMRU)

Sample preparation, extraction and fractionation were conducted at SMRU, whilst all instrumental analyses were carried out by MAFF. The basic procedures are outlined in Section 3.3 above. The organochlorine data

are given in Tables 7 - 10. Tables 7 - 9 relate to analyses of blubber, and Table 10 to analyses of stomach contents from a particular group of common dolphins. Table 7 presents data for total chlorobiphenyls (both the sum of the 7 ICES congeners, and the sum of the 25 CBs determined in this study; see Table 1 for the identity of the 25 congeners analysed) and for a range of organochlorine compounds including pesticides. Priority in this study was given to analysis of chlorobiphenyls, and in many cases the fraction containing the majority of the pesticides was not analysed. Dashes in the relevant columns indicate where data are unavailable for this reason — where analyses were conducted but the concentration was below the limit of detection 'nd' (not detected) is displayed.

The ICES 7 congeners were selected by the International Council for the Exploration of the Sea as a basic list for monitoring purposes (Anon., 1986) and comprise CBs 28, 52, 101, 118, 138, 153 and 180. Tables 8 and 9 present the data for the 25 individual CB congeners, in their order of elution from a low-polarity (5% phenylmethyl silicone) stationary phase. In all these tables the data are reported in units of mg kg⁻¹ wet weight, but the content of hexane-extractable lipid is also given so that concentrations can be recalculated on a lipid basis if required, and the reference numbers are given for each animal in order to provide a link to Tables 2 and 3.

5. SUMMARY AND CONCLUSION

Thijs Kuiken (IOZ) and Robin J. Law (MAFF)

Through this project detailed postmortem examinations were carried out on more than 300 marine mammal carcasses from around the coast of the UK. As a result of the co-operative effort of a number of institutes, it was possible to attribute the cause of a mass mortality of common dolphins around the south-west of England to accidental capture in fishing gear (Kuiken *et al.*, 1994(a)). Recommendations were made on how to assess the importance of this cause of death for the common dolphin population in these waters. Accidental capture in fishing gear was also the single most important cause of death for harbour porpoises from British waters (Kuiken *et al.*, 1994(b)). The first cases of morbillivirus infection in harbour porpoises in Great Britain were diagnosed (Kennedy *et al.*, 1992), and several marine mammal species found stranded on the coast of England and Wales had serological evidence of infection with a morbillivirus (Visser *et al.*, 1993) and a newly discovered virus with a rhabdovirus morphology (Osterhaus *et al.*, 1993). Bacterial and parasitic infections were found regularly in harbour porpoises and common dolphins, and were occasionally fatal (Kuiken *et al.*, 1994(a) and (b)).

Table 2. Basic information for animals other than harbour porpoises

ReferenceNo.	Species	Sex	Age (yrs)	Length (cm)	Found	Location	County / Region
LAW-19	H. grypus	M	-	-	22/10/88	Donna Nook	Lincolnshire
LAW-20	H. grypus	M	-	-	??/10/88	Cardigan Bay	Dyfed
LAW-21	H. grypus	-	-	-	??/10/88	Cardigan Bay	Dyfed
LAW-22	H. grypus	M	-	215	06/12/88	Ceibwr Bay	Dyfed
LAW-23	H. grypus	F	< 1	-	23/12/88	River Tees	Cleveland
LAW-24	H. grypus	F	-	100	10/01/89	Portaferry	Co.Down
LAW-25	H. grypus	F	-	99	05/02/89	Portrush	Co. Antrim
LAW-26	H. grypus	M	-	189	19/05/89	West Kirby, Wirral	Merseyside
LAW-27	H. grypus	F	-	-	01/06/89	Portrush	Co. Antrim
LAW-28	H. grypus	M	-	212	02/06/89	West Kirby, Wirral	Merseyside
LAW-29	H. grypus	M	-	188	27/06/89	Mostyn	Clwyd
LAW-30	H. grypus	F	-	168	01/07/89	River Dee	Clwyd / Merseyside
LAW-31	H. grypus	M	-	209	10/07/89	West Kirby, Wirral	Merseyside
LAW-32	H. grypus	F	-	183	28/09/89	New Quay	Dyfed
LAW-33	H. grypus	M	-	-	18/10/89	Portstewart	Co. Antrim
LAW-34	H. grypus	M	-	97	25/01/90	Llandudno	Gwynedd
LAW-35	H. grypus	F	-	145	11/06/90	Dove Point, Hoylake	Merseyside
LAW-36	H. grypus	F	-	-	09/07/90	Fomby	Merseyside
LAW-37	H. grypus	M	-	182	10/07/90	Fomby	Merseyside
LAW-38	H. grypus	M	-	115	??/07/90	Port Erin	Isle of Man
LAW-39	H. grypus	M	-	208	02/10/90	West Kirby, Wirral	Merseyside
SS1990/3	H. grypus	F	-	183	08/11/90	New Quay	Dyfed
SS1990/7	H. grypus	F	-	165	23/11/90	Druidston Haven	Dyfed
SS1990/11	H. grypus	F	-	102	02/12/90	St Ives	Cornwall
SS1990/12A	H. grypus	M	-	116	10/12/90	Beadnell	Northumberland
SS1990/12B	H. grypus	F	-	108	10/12/90	Beadnell	Northumberland
SS1990/16	H. grypus	F	2	-	24/12/90	Gweek (seal sanctuary)	Cornwall
SS1991/1A	H. grypus	M	-	109	01/01/91	Hayle Towan	Cornwall
SS1991/1B	H. grypus	M	-	107	04/01/91	St.Michael's Mount	Cornwall
SS1991/9	H. grypus	F	-	96	21/01/91	Unknown	Devon
SS1991/23	H. grypus	F	-	160	09/04/91	Blackpool	Lancashire
SS1991/27	H. grypus	M	-	178	17/05/91	Fomby	Merseyside
SS1991/28	H. grypus	M	-	206	22/05/91	Wirral	Cheshire
SS1991/30A	H. grypus	M	-	155	18/06/91	Caldy	Merseyside
SS1991/41	H. grypus	F	-	191	27/08/91	Hoylake	Merseyside
SS1991/50	H. grypus	F	-	92	??/10/91	Ramsey	Dyfed
SS1991/55	H. grypus	M	-	107	12/11/91	Borth	Dyfed
SS1991/58	H. grypus	M	-	180	15/11/91	Llangranog	Dyfed
SS1991/73	H. grypus	M	-	100	20/12/91	Llandanwg	Gwynedd
SS1992/9	H. grypus	F	-	178	25/03/92	Llanon	Dyfed
LAW-40	P. vitulina	M	5	-	25/08/88	Blakeney Point	Norfolk
LAW-41	P. vitulina	M	2	-	25/08/88	Blakeney Point	Norfolk
LAW-42	P. vitulina	F	1	-	25/08/88	Blakeney Point	Norfolk
LAW-43	P. vitulina	M	1	-	25/08/88	Blakeney Point	Norfolk
LAW-44	P. vitulina	M	6	152	31/08/88	River Humber	Humberside
LAW-45	P. vitulina	F	-	130	12/09/88	Blakeney Point	Norfolk
LAW-46	P. vitulina	F	4	-	12/09/88	Blakeney Point	Norfolk
LAW-47	P. vitulina	M	2	122	13/09/88	Blakeney Point	Norfolk
LAW-48	P. vitulina	F	14	131	14/09/88	Blakeney Point	Norfolk
LAW-49	P. vitulina	F	2	120	15/09/88	Blakeney Point	Norfolk
LAW-50	P. vitulina	F	8	142	16/09/88	Blakeney Point	Norfolk
LAW-51	P. vitulina	M	-	-	16/09/88	Strangford	Co.Down
LAW-52	P. vitulina	M	6	149	18/09/88	Blakeney Point	Norfolk
LAW-53	P. vitulina	F	-	-	23/09/88	Blakeney Point	Norfolk
LAW-54	P. vitulina	M	-	-	26/09/88	Newcastle	Co.Down
LAW-55	P. vitulina	M	-	-	29/09/88	Unknown	Co.Down
LAW-56	P. vitulina	M	-	-	29/09/88	Unknown	Co.Down
LAW-57	P. vitulina	M	-	105	03/10/88	Ballykinlar	Co.Down
LAW-58	P. vitulina	M	-	-	06/10/88	Annalong	Co.Down
LAW-59	P. vitulina	M	-	-	17/10/88	Unknown	Co.Down
LAW-60	P. vitulina	F	-	147	24/10/88	Whiteabbey	Co. Antrim
LAW-61	P. vitulina	M	-	100	26/10/88	Whitehead	Co. Antrim
LAW-62	P. vitulina	F	-	-	26/10/88	Blakeney Point	Norfolk
LAW-63	P. vitulina	F	-	-	04/11/88	Blakeney Point	Norfolk
LAW-64	P. vitulina	F	-	165	09/11/88	Kilkeel	Co.Down
LAW-65	P. vitulina	F	-	144	22/11/88	Rathlin Island	Co. Antrim
LAW-66	P. vitulina	M	< 1	-	??/12/88	River Tees	Cleveland
LAW-67	P. vitulina	F	-	70	05/05/89	Unknown	Co.Down
LAW-68	P. vitulina	M	< 1	-	15/07/89	River Tees	Cleveland
LAW-69	P. vitulina	M	-	-	18/10/89	Portaferry	Co.Down
SW1991/2	B. acutorostrata	F	-	-	02/01/91	Carmarthen Bay, Pembrey	Dyfed

Table 2. (continued)

Reference No.	Species	Sex	Age (yrs)	Length (cm)	Found	Location	County / Region
SMRU90-73	<i>D. delphis</i>	-	-	-	??/??/90	Unknown	Unknown
IC1990/4	<i>D. delphis</i>	M	-	-	??/10/90	Seaton	Cornwall
IC1991/1A	<i>D. delphis</i>	F	-	194	07/01/91	Porthcawl	Mid Glamorgan
IC1991/1B	<i>D. delphis</i>	F	foetus	39	07/01/91	Porthcawl	Mid Glamorgan
SW1990/5A	<i>D. delphis</i>	F	-	167	19/01/90	Langness	Isle of Man
SW1990/20A	<i>D. delphis</i>	-	-	-	??/??/90	Unknown	Unknown
SW1990/26	<i>D. delphis</i>	F	-	162	05/03/90	Carmarthen Bay	Dyfed
SW1990/45	<i>D. delphis</i>	F	-	168	21/05/90	Swansea	West Glamorgan
SW1990/46	<i>D. delphis</i>	M	-	-	22/05/90	Lambeth Bridge	Greater London
SW1990/47	<i>D. delphis</i>	F	-	190	23/05/90	Wapping	Greater London
SW1990/68	<i>D. delphis</i>	F	17	194	01/09/90	Stoke Beach	Devon
SW1990/78	<i>D. delphis</i>	M	18	220	26/09/90	Seaton Beach, Downderry	Cornwall
SW1990/81	<i>D. delphis</i>	F	-	162	02/10/90	Caer-fai Bay	Dyfed
SW1990/87	<i>D. delphis</i>	M	?5	176	16/10/90	Caerthilian Cove	Cornwall
SW1990/88	<i>D. delphis</i>	-	?6	197	22/10/90	Truro	Cornwall
SW1990/90A	<i>D. delphis</i>	-	9	-	24/10/90	Helford River	Cornwall
SW1990/97	<i>D. delphis</i>	M	1	132	16/11/90	Porthmeor	Cornwall
SW1990/101(1)	<i>D. delphis</i>	M	?13	206	25/11/90	Bexhill on Sea	East Sussex
SW1990/101(2)	<i>D. delphis</i>	F	4-5	181	25/11/90	Bexhill on Sea	East Sussex
SW1990/103	<i>D. delphis</i>	F	6	167	26/11/90	Eastbourne	East Sussex
SW1990/104	<i>D. delphis</i>	F	?9	178	26/11/90	Cuckmere Haven	East Sussex
SW1990/108	<i>D. delphis</i>	M	14	212	07/12/90	Gorran Haven Beach	Cornwall
SW1991/7	<i>D. delphis</i>	F	-	226	10/01/91	Banjo Groyne, Brighton	East Sussex
SW1991/10	<i>D. delphis</i>	M	-	174	09/01/91	Harlech	Gwynedd
SW1991/37	<i>D. delphis</i>	F	3	239?	16/04/91	Aberavon Sands, Port Talbot	West Glamorgan
SW1991/40	<i>D. delphis</i>	F	-	187	12/04/91	Seaton	Cornwall
SW1991/46	<i>D. delphis</i>	M	-	210	28/04/91	Lansallos	Cornwall
SW1991/83	<i>D. delphis</i>	F	?10	198	18/07/91	Cemaes Head	Dyfed
SW1991/129	<i>D. delphis</i>	M	<1	125	06/10/91	Lawrenny Quay	Dyfed
SW1991/137	<i>D. delphis</i>	F	20+	205	27/10/91	Swanage	Dorset
SW1992/1	<i>D. delphis</i>	F	-	191	02/01/92	Looe Beach	Cornwall
SW1992/2	<i>D. delphis</i>	M	-	205	07/01/92	Penzance	Cornwall
SW1992/3	<i>D. delphis</i>	F	-	-	08/01/92	Porthleven Harbour	Cornwall
SW1992/4	<i>D. delphis</i>	M	-	195	14/01/92	Praa Sands	Cornwall
SW1992/10	<i>D. delphis</i>	F	-	183	02/02/92	Penzance	Cornwall
SW1992/11	<i>D. delphis</i>	F	-	184	02/02/92	Porthtowan	Cornwall
SW1992/12	<i>D. delphis</i>	M	-	220	02/02/92	St Keverne	Cornwall
SW1992/14 (1)	<i>D. delphis</i>	F	-	-	07/02/92	Falmouth	Cornwall
SW1992/14 (2)	<i>D. delphis</i>	F	-	-	07/02/92	Falmouth	Cornwall
SW1992/19 (1)	<i>D. delphis</i>	F	-	171	13/02/92	Bigbury	Devon
SW1992/19 (2)	<i>D. delphis</i>	F	-	-	13/02/92	Bigbury	Devon
SW1992/20	<i>D. delphis</i>	F	-	204	13/02/92	Kingsbridge	Devon
SW1992/172 (1)	<i>D. delphis</i>	F	28	205	03/08/92	Newport	Dyfed
SW1992/172 (2)	<i>D. delphis</i>	F	2 - 3	150	03/08/92	Newport	Dyfed
SW1990/70	<i>G. griseus</i>	F	-	161	03/09/90	Porthmadog	Gwynedd
SW1990/88A	<i>G. melas</i>	M	-	190	20/10/90	Maenporth Beach	Cornwall
SW1991/55	<i>G. melas</i>	M	-	588	20/05/91	Redcar	Cleveland
LAW-70	<i>L. acutus</i>	-	-	-	02/03/89	Unknown	Co. Sligo
SW1989/88B	<i>L. acutus</i>	M	-	-	03/11/89	Unknown	Shetland
SW1990/114	<i>L. acutus</i>	M	<1	158	27/12/90	Beadnell	Northumberland
SW1991/9	<i>L. acutus</i>	F	10	217	11/01/91	Saunton Sands	Devon
SW1989/49	<i>L. albirostris</i>	F	-	255	28/06/89	West Kirby, Wirral	Merseyside
SW1991/108	<i>L. albirostris</i>	F	-	217	04/09/91	Cayton Bay, Osgodby	North Yorkshire
SW1991/126	<i>L. albirostris</i>	M	-	249	30/09/91	Withernsea	Humberside
SW1990/4	<i>S. coeruleoalba</i>	M	-	186	22/01/90	Carmarthen Bay	Dyfed
SW1990/20	<i>S. coeruleoalba</i>	M	-	185	14/02/90	Aberstwyth	Dyfed
SW1990/57	<i>S. coeruleoalba</i>	M	-	166	09/07/90	Brighton Marina	East Sussex
SW1991/11	<i>S. coeruleoalba</i>	M	6	187	13/01/91	Mill Haven	Dyfed
SW1991/18	<i>S. coeruleoalba</i>	M	3	157	06/02/91	Aldeburgh	Suffolk
SW1991/26	<i>S. coeruleoalba</i>	M	-	190	20/02/91	Talacre	Clwyd
SW1991/122	<i>S. coeruleoalba</i>	M	-	235	27/09/91	Port Soif	Guernsey
LAW-71	<i>T. truncatus</i>	F	-	271	25/06/89	Tenby	Dyfed
SW1989/47	<i>T. truncatus</i>	-	-	217	26/06/89	Pen-y-Bryn, New Quay	Dyfed
SW1991/42	<i>T. truncatus</i>	M	23	342	17/04/91	Upper Borth	Dyfed
SW1991/115	<i>T. truncatus</i>	F	-	154	13/09/91	Beaumaris	Gwynedd
SW1991/130	<i>T. truncatus</i>	F	-	322	07/10/91	Bognor Regis	West Sussex
SW1992/25	<i>T. truncatus</i>	M	-	183	19/02/92	Harlech	Gwynedd

Table 3. Basic information and probable cause of death for harbour porpoises

ReferenceNo.	Species	Sex	Age (yrs)	Length (cm)	Found	Location	County / Region	Probable Cause of Death
IC1990/1	P. phocoena	F	1	128	20/04/90	Usk mouth	Gwent	by-catch
IC1990/2	P. phocoena	F	1	117	20/08/90	Puffin Island	Gwynedd	by-catch
IC1990/3	P. phocoena	F	-	118	15/10/90	Caernarfon	Gwynedd	starvation
IC1990/4	P. phocoena	M	-	108	??/??/90	Caernarfon Bay	Gwynedd	by-catch
IC1991/2	P. phocoena	M	0	78	15/07/91	Anglesey	Gwynedd	starvation (neonate)
91L-2947	P. phocoena	M	-	84	22/06/89	Swansea Bay	West Glamorgan	physical trauma
SMRU90-01	P. phocoena	M	10	145	06/12/89	Unknown	Shetland	physical trauma, by-catch*
SMRU90-04	P. phocoena	M	1	119	??/??/??	Teesside	Cleveland	no post-mortem done
SMRU90-02	P. phocoena	F	6	157	02/12/89	Unknown	Shetland	by-catch
SMRU90-03	P. phocoena	M	11	143	06/12/89	Unknown	Shetland	by-catch
SMRU90-05	P. phocoena	M	4	141	13/10/89	Unknown	Shetland	by-catch
SMRU90-06	P. phocoena	F	2	133	30/08/89	Unknown	Shetland	by-catch
SMRU90-07	P. phocoena	F	5	150	02/10/89	Unknown	Shetland	by-catch
SMRU90-08	P. phocoena	F	4	152	20/10/89	Unknown	Shetland	by-catch
SMRU90-09	P. phocoena	M	1	114	24/08/89	Unknown	Shetland	by-catch
SMRU90-10	P. phocoena	M	3	142	24/08/89	Unknown	Shetland	by-catch
SMRU90-11	P. phocoena	F	2	147	23/08/89	Unknown	Shetland	by-catch
SMRU90-12	P. phocoena	F	10	153	24/08/89	Unknown	Shetland	by-catch
SMRU90-13	P. phocoena	F	3	148	27/07/89	Unknown	Shetland	by-catch
SMRU90-14	P. phocoena	F	1	123	14/10/89	Unknown	Shetland	by-catch
SMRU90-15	P. phocoena	M	9	139	01/11/89	Unknown	Shetland	by-catch
SMRU90-16	P. phocoena	M	5	145	28/07/89	Unknown	Shetland	by-catch
SMRU90-17	P. phocoena	M	5	142	20/07/89	Unknown	Shetland	by-catch
SMRU90-18	P. phocoena	M	8	140	05/08/89	Unknown	Shetland	no post-mortem done
SMRU90-19	P. phocoena	M	9	146	11/07/89	Unknown	Shetland	by-catch
SW1989/43A	P. phocoena	F	-	171	19/06/89	Hoylelake	Merseyside	not established
SW1989/51	P. phocoena	F	-	75	03/07/89	Swansea Bay	West Glamorgan	not established
SW1989/89	P. phocoena	F	-	173	05/11/89	Newport	Dyfed	parasitic pneumonia
SMRU90-51	P. phocoena	F	3	157	??/??/??	River Blackwater	Essex	parasitic pneumonia
SMRU90-55	P. phocoena	M	1	113	??/??/??	River Blackwater	Essex	<i>Streptococcus</i> pneumonia
SMRU90-56	P. phocoena	F	4	156	13/05/90	Unknown	Western Isles	not established
SMRU90-58	P. phocoena	M	3	151	07/06/90	Unknown	Shetland	by-catch
SMRU90-59	P. phocoena	M	2	125	??/??/90	Unknown	Shetland	by-catch
SMRU90-60	P. phocoena	M	2	143	??/??/90	Unknown	Shetland	by-catch
SMRU90-61	P. phocoena	M	4	137	19/07/90	Dysart	Fife	not established
SMRU90-72A	P. phocoena	F	-	-	12/09/90	Mablethorpe	Lincolnshire	no post-mortem done
SMRU90-75	P. phocoena	-	-	-	??/??/??	Clyde Estuary		no post-mortem done
SMRU90-75A	P. phocoena	-	-	-	01/12/88	Unknown	Scotland	no post-mortem done
SMRU90-76	P. phocoena	M	-	-	??/??/??	Holkham & Wells	Norfolk	no post-mortem done
SW1990/31	P. phocoena	F	-	133	25/03/90	St David's	Dyfed	peritonitis, gastritis (pyloric section)
SW1990/39	P. phocoena	F	1	105 §	08/04/90	Tenby	Dyfed	bacterial pneumonia
SW1990/50	P. phocoena	F	-	138	20/06/90	Carmarthen Bay	Dyfed	<i>Pasteurella</i> septicaemia
SW1990/50A	P. phocoena	M	0	73	24/06/90	Ynyslas, Borth	Dyfed	physical trauma
SW1990/51(1)	P. phocoena	F	-	165	28/06/90	Pembrey	Dyfed	dystocia
SW1990/51(2)	P. phocoena	M	0	80	28/06/90	Pembrey	Dyfed	dystocia
SW1990/54	P. phocoena	M	0	87	02/07/90	Newport	Gwent	starvation
SW1990/65	P. phocoena	M	8	148	14/08/90	Aberdyfi	Gwynedd	bacterial pneumonia
SW1990/66	P. phocoena	M	-	-	18/08/90	Redruth	Cornwall	no post-mortem done
SW1990/71	P. phocoena	F	4	146	14/08/90	Aberdeen	Grampian	by-catch
SW1990/72	P. phocoena	F	-	106	06/09/90	Abersoch	Gwynedd	suffocation
SW1990/72A	P. phocoena	M	-	139	05/09/90	Pembrey	West Glamorgan	parasitic pneumonia
SW1990/73	P. phocoena	M	0	79	05/09/90	Unknown	Dyfed	<i>Clostridium</i> myositis
SW1990/77A	P. phocoena	M	-	142	24/09/90	Porthcawl	Mid Glamorgan	not established
SW1990/77B	P. phocoena	M	-	138	25/09/90	Llanelli	Dyfed	by-catch
SW1990/79	P. phocoena	M	1	125	27/09/90	Sunderland	Tyne & Wear	by-catch
SW1990/84	P. phocoena	F	6	150	07/10/90	Bridlington	Humberside	parasitic pneumonia
SW1990/92	P. phocoena	F	-	180	30/10/90	Ainsdale	Merseyside	adenocarcinoma
SW1990/94	P. phocoena	F	11	151	07/11/90	Sea Palling Beach	Norfolk	pneumonia
SW1990/96	P. phocoena	M	4	135	10/11/90	Penzance	Cornwall	parasitic bronchitis
SW1990/98	P. phocoena	F	-	125	18/11/90	Dale	Dyfed	parasitic & bacterial pneumonia
SW1990/100	P. phocoena	F	-	125	21/11/90	Shell Island	Gwynedd	mycotic pneumonia
SW1990/100A	P. phocoena	M	0	102	??/??/??	Portslade	East Sussex	starvation
SW1990/106	P. phocoena	M	1	112	29/11/90	Isle of Sheppey	Kent	morbillivirus infection
SW1990/107	P. phocoena	F	-	176	05/12/90	Druidston	Dyfed	parasitic & bacterial pneumonia
SW1990/109	P. phocoena	F	0	-	??/12/90	Sennen Cove	Cornwall	suspected by-catch
SW1990/111	P. phocoena	M	-	137	19/12/90	Borth	Dyfed	suspected by-catch
SMRU91-06	P. phocoena	F	8	145	26/07/91	Isle of Noss	Shetland	not established
SMRU91-08	P. phocoena	F	4	158	08/10/91	Weisdale Voe	Shetland	neoplasia
91L-2937	P. phocoena	F	0	117	12/09/91	Dinas, Caernarfon	Gwynedd	generalised <i>Streptococcus lactis</i> infection

Table 3. (continued)

Reference No.	Species	Sex	Age (yrs)	Length (cm)	Found	Location	County / Region	Probable Cause of Death
91L-2936	<i>P. phocoena</i>	M	-	109	04/10/91	Morfa, Harlech	Gwynedd	enteritis
SMRU91-15	<i>P. phocoena</i>	M	0	107	??/??/??	Unknown	Isle of Man	by-catch
SMRU91-07	<i>P. phocoena</i>	F	1	143	30/09/91	Bard Head, Bressay	Shetland	by-catch
SMRU91-16	<i>P. phocoena</i>	M	-	-	17/10/91	Port Erin	Isle of Man	not established
SW1991/14	<i>P. phocoena</i>	F	14	189	??/01/91	Wherrytown	Cornwall	starvation
SW1991/17	<i>P. phocoena</i>	M	8	141	29/01/91	Joss Bay	Kent	pneumonia
SW1991/17A	<i>P. phocoena</i>	F	4	137	05/02/91	Shanklin	Isle of Wight	parasitic pneumonia
SW1991/19A	<i>P. phocoena</i>	M	5	150	12/02/91	Skinningrove	Cleveland	parasitic lung haemorrhage
SW1991/19B	<i>P. phocoena</i>	F	2	131	??/02/91	Porthmeor, St Ives	Cornwall	suffocation
SW1991/20	<i>P. phocoena</i>	F	1	132	07/02/91	Southwold Beach	Suffolk	parasitic pneumonia
SW1991/22	<i>P. phocoena</i>	M	11	152	14/02/91	South Shields	Tyne & Wear	parasitic stomach haemorrhage
SW1991/23	<i>P. phocoena</i>	M	0	103	18/02/91	Bridlington Bay	Humberside	physical trauma
SW1991/24	<i>P. phocoena</i>	M	7	143	16/02/91	Bexhill	East Sussex	unknown
SW1991/28	<i>P. phocoena</i>	F	3	132	07/03/91	Hornsea	Humberside	by-catch
SW1991/29	<i>P. phocoena</i>	F	0	110	11/03/91	Bridlington	Humberside	physical trauma
SW1991/30	<i>P. phocoena</i>	M	1	94	09/03/91	Warkworth	Northumberland	unknown
SW1991/31	<i>P. phocoena</i>	M	2	111	13/03/91	Seahouses	Northumberland	inhalation pneumonia
SW1991/32A	<i>P. phocoena</i>	M	2	151	17/03/91	Carne Beach, Veryan	Cornwall	physical trauma
SW1991/35	<i>P. phocoena</i>	M	0	100	27/03/91	South Shields	Tyne & Wear	starvation
SW1991/36	<i>P. phocoena</i>	F	14	152	02/04/91	Fort Victoria	Isle of Wight	by-catch
SW1991/43	<i>P. phocoena</i>	F	9	148	20/04/91	Filey Brigg	North Yorkshire	parasitic pneumonia
SW1991/48	<i>P. phocoena</i>	F	-	120	25/04/91	Mablethorpe	Lincolnshire	<i>Erwinia</i> meningo-encephalitis
SW1991/54	<i>P. phocoena</i>	F	3	128	13/05/91	Foulness Point	Essex	parasitic stomach haemorrhage
SW1991/59	<i>P. phocoena</i>	F	0	82	07/06/91	Gorleston Beach	Norfolk	starvation
SW1991/61	<i>P. phocoena</i>	F	0	72	10/06/91	Ferryside	Dyfed	starvation
SW1991/63	<i>P. phocoena</i>	F	0	76	12/06/91	Skegness	Lincolnshire	not established
SW1991/70	<i>P. phocoena</i>	M	1	102	22/06/91	Blyth	Northumberland	not established
SW1991/71	<i>P. phocoena</i>	M	-	127	24/06/91	Anglesey	Gwynedd	enteritis
SW1991/73	<i>P. phocoena</i>	M	0	88	26/06/91	Hunstanton	Norfolk	starvation
SW1991/74	<i>P. phocoena</i>	M	0	78	29/06/91	Amroth Beach, Tenby	Dyfed	not established
SW1991/84	<i>P. phocoena</i>	F	0	72	17/07/91	Harlech	Gwynedd	starvation
SW1991/95	<i>P. phocoena</i>	F	-	140	30/07/91	Great Yarmouth	Norfolk	not established
SW1991/96	<i>P. phocoena</i>	M	0	87	01/08/91	Lowestoft	Suffolk	starvation
SW1991/100	<i>P. phocoena</i>	M	0	60	12/08/91	Southernness Point	Dumfries & Galloway	physical trauma
SW1991/104	<i>P. phocoena</i>	M	12	142	29/08/91	Hornsea	Humberside	physical trauma
SW1991/109	<i>P. phocoena</i>	F	1	114	06/09/91	Reighton Sands	North Yorkshire	not established
SW1991/111	<i>P. phocoena</i>	F	1	134	05/09/91	Robin Hood's Bay	North Yorkshire	not established
SW1991/112	<i>P. phocoena</i>	M	10	136	06/09/91	Hornsea	Humberside	by-catch
SW1991/116	<i>P. phocoena</i>	F	5	152	16/09/91	Clacton on Sea	Essex	not established
SW1991/117A	<i>P. phocoena</i>	M	0	97	01/08/91	Holland on Sea	Essex	starvation
SW1991/120	<i>P. phocoena</i>	F	0	117	23/09/91	Burry Port	Dyfed	bacterial pneumonia
SW1991/124	<i>P. phocoena</i>	F	-	134	01/10/91	Skomer Island	Dyfed	physical trauma
SW1991/128	<i>P. phocoena</i>	M	3	130	02/10/91	Saltburn by the Sea	Cleveland	pneumonia
SW1991/135	<i>P. phocoena</i>	F	0	87	21/10/91	Blackpool	Lancashire	starvation
SW1991/136	<i>P. phocoena</i>	M	-	144	22/10/91	Cardigan Island	Dyfed	parasitic pneumonia
SW1991/142	<i>P. phocoena</i>	F	-	118	13/11/91	Towyn	Clwyd	physical trauma
SW1992/7	<i>P. phocoena</i>	F	-	131	17/01/92	Borth	Dyfed	suffocation
SW1992/112	<i>P. phocoena</i>	M	-	133	15/05/92	Tywyn	Gwynedd	by-catch
SW1992/170	<i>P. phocoena</i>	M	6	157	29/07/92	Tywyn	Gwynedd	neck fracture
SW1992/175	<i>P. phocoena</i>	M	4	140	03/08/92	Pembrey	Dyfed	not established
LAW-01	<i>P. phocoena</i>	F	-	140	??/??/88	Unknown	County Down	morbillivirus infection
LAW-02	<i>P. phocoena</i>	F	-	135	??/??/88	Unknown	County Down	morbillivirus infection
LAW-03	<i>P. phocoena</i>	M	0	70	??/06/88	Islay	Western Isles	not established
LAW-04	<i>P. phocoena</i>	F	-	86	01/06/88	Derbyhaven Bay	Isle of Man	not established
LAW-05	<i>P. phocoena</i>	-	-	-	??/09/88	Cardigan Bay	Dyfed	not established
LAW-06	<i>P. phocoena</i>	-	-	-	24/09/88	Aberaeron	Dyfed	not established
LAW-07	<i>P. phocoena</i>	M	-	-	27/10/88	Fleshwick Bay	Isle of Man	not established
LAW-08	<i>P. phocoena</i>	M	-	147	09/11/88	Kilkeel	County Down	morbillivirus infection
LAW-09	<i>P. phocoena</i>	-	-	109	19/12/88	Crawfordsburn	County Down	morbillivirus infection
LAW-10	<i>P. phocoena</i>	F	-	163	04/01/89	Castlerock	County Antrim	liver abscess ?
LAW-11	<i>P. phocoena</i>	F	-	125	17/02/89	Folkestone	Kent	by-catch*
LAW-12	<i>P. phocoena</i>	M	-	135	17/02/89	Folkestone	Kent	by-catch*
LAW-13	<i>P. phocoena</i>	F	-	135	02/03/89	off Hanna Voe	Shetland	not established
LAW-14	<i>P. phocoena</i>	F	-	-	07/03/89	Unknown	Shetland	by-catch*
LAW-15	<i>P. phocoena</i>	-	-	72	26/06/89	Borth	Dyfed	not established
LAW-16	<i>P. phocoena</i>	-	-	71	27/06/89	Cei Bach	Dyfed	not established
LAW-17	<i>P. phocoena</i>	M	-	102	08/08/89	Whitby	North Yorkshire	by-catch*
LAW-18	<i>P. phocoena</i>	F	-	79	??/08/90	Unknown	South Wales	not established

§ approximate: most of head absent

When the term 'by-catch*' is given in the probable cause of death column this means that the animal was found entangled in fishing nets, but no post-mortem was carried out. This distinguishes it from 'by-catch' where post-mortem investigations were used to establish the cause of death

Table 4. Concentrations of trace metals in liver samples (mg kg⁻¹ wet weight)

Reference No.	%Dry Matter	Cr	Ni	Cu	Zn	As	Se	Cd	Hg	Pb
H. grypus										
LAW-19	30.5	<0.5	<0.5	22	25	-	-	0.06	110	<0.6
LAW-20	30.1	<0.5	<0.5	26	80	-	-	<0.06	1.7	<0.6
LAW-21	26.5	<0.6	<0.6	9.1	88	-	-	<0.07	2.5	<0.7
LAW-22	26.2	<0.5	<0.5	7.2	68	-	-	0.12	45	<0.6
LAW-23	33.7	<0.5	<0.5	3.8	60	-	-	<0.06	0.26	<0.6
LAW-24	36.1	0.6	<0.5	13	36	-	-	0.28	3.9	<0.6
LAW-25	30.9	<0.5	<0.5	14	63	-	-	0.16	3.5	<0.6
LAW-26	28.7	<0.5	<0.5	18	53	-	-	0.16	430	1
LAW-27	30.9	<0.5	<0.5	7.8	61	-	-	0.06	7.9	<0.6
LAW-28	31.1	<0.5	<0.5	22	40	-	-	0.19	210	1.8
LAW-29	26.4	<0.5	<0.5	14	89	-	-	0.2	370	1.4
LAW-30	25	<0.5	<0.5	7.7	89	-	-	1.2	12	0.6
LAW-31	25	0.8	<0.5	14	43	-	-	<0.07	110	1
LAW-32	23.9	2	2.1	29	110	-	-	0.22	210	0.15
LAW-33	25.6	<0.5	<0.5	39	61	-	-	<0.06	11	0.53
LAW-34	28.9	0.5	1.2	6.1	24	-	-	<0.06	1.5	0.56
LAW-35	25.4	<0.5	<0.5	15	28	-	-	<0.06	31	1.7
LAW-36	26.8	<0.5	<0.5	12	67	-	-	0.97	20	0.73
LAW-37	26.8	<0.5	<0.5	14	37	-	-	0.2	120	1.3
LAW-38	29.3	<0.5	<0.5	7.9	28	-	-	0.33	220	7
LAW-39	32.9	<0.5	<0.5	28	85	-	-	0.79	280	0.95
SS1990/3	30.9	<0.6	<0.6	48	51	-	45	1.9	131	0.17
SS1990/7	20.9	<0.56	<0.56	4.1	19	-	5.7	2	6.1	<0.03
SS1990/11	22.8	<0.54	<0.54	8.7	55	-	0.9	<0.06	1.2	<0.03
SS1990/12A	20.1	<0.54	<0.54	7.6	47	-	0.8	<0.07	0.27	<0.03
SS1990/12B	25.5	<0.55	<0.55	11	53	-	1.3	<0.07	0.47	<0.03
SS1990/16	26.1	0.66	<0.51	18	57	-	2.1	0.1	2	<0.03
SS1991/1A	26.4	<0.55	<0.5	16	53	-	0.7	<0.06	2.2	0.22
SS1991/1B	24.3	<0.55	<0.55	12	46	-	1.2	<0.07	0.78	0.07
SS1991/9	27	<0.65	<0.65	15	71	-	1.2	0.27	0.07	0.06
SS1991/23	24.6	0.6	<0.5	5.2	77	-	-	0.64	170	1.9
SS1991/27	26.5	<0.5	<0.5	2.2	22	-	-	0.41	14	0.38
SS1991/28	25.6	<0.44	<0.44	9.9	60	-	68	<0.05	52	0.49
SS1991/30A	25.5	0.63	<0.46	3.2	17	-	30	0.33	80	0.43
SS1991/41	26.3	<0.51	<0.51	18	36	-	123	0.14	301	1.7
SS1991/50	23.5	<0.51	<0.51	40	55	-	2.4	<0.06	1.3	0.11
SS1991/55	19.5	<0.5	<0.5	1.9	29	-	0.25	<0.06	0.22	<0.03
SS1991/58	29.2	<0.53	<0.53	18	91	-	29	0.18	65	0.07
SS1991/73	20.5	1	<0.51	17	113	-	0.82	<0.06	2.6	0.06
SS1992/9	26.1	0.67	<0.53	3.6	33	-	37	0.15	65	0.05
P. vitulina										
LAW-40	26.4	<0.6	<0.6	6.2	42	-	-	0.12	57	<0.7
LAW-41	30.3	<0.6	<0.6	12	41	-	-	<0.07	16	<0.7
LAW-42	31.6	<0.6	<0.6	5.8	48	-	-	<0.07	6.4	<0.7
LAW-43	28.9	<0.5	<0.5	6.9	41	-	-	<0.06	6.2	<0.6
LAW-44	26.6	<0.7	<0.7	7.4	39	-	-	0.1	110	<0.8
LAW-45	27.5	<0.5	<0.5	7.4	50	-	-	0.18	100	<0.6
LAW-46	28.6	<0.5	<0.5	6.2	43	-	-	0.14	36	<0.7
LAW-47	30	<0.6	<0.6	9.3	44	-	-	0.11	9.6	<0.7
LAW-48	29	<0.7	<0.7	9	57	-	-	0.3	98	<0.8
LAW-49	29.2	<0.5	<0.5	6	66	-	-	0.11	21	<0.6
LAW-50	28.1	<0.6	<0.6	6.6	47	-	-	0.2	59	<0.7
LAW-51	29.2	1	<0.5	10	49	-	-	0.29	49	<0.6
LAW-52	29.1	<0.6	<0.6	6.7	49	-	-	0.15	9.6	<0.7
LAW-53	29.3	<0.6	<0.6	5.2	32	-	-	0.17	81	<0.7
LAW-54	27.6	0.9	<0.5	17	49	-	-	0.94	170	<0.6
LAW-55	27.7	<0.5	<0.5	21	58	-	-	0.2	45	<0.6
LAW-56	26.8	<0.6	<0.5	15	72	-	-	0.12	66	<0.6
LAW-57	26	<0.5	<0.5	7.8	47	-	-	<0.06	6.3	<0.6
LAW-58	27.4	<0.5	<0.5	20	48	-	-	0.79	160	<0.6
LAW-59	25.4	<0.6	<0.5	17	48	-	-	<0.06	7.5	<0.6
LAW-60	28.3	<0.5	<0.5	13	55	-	-	0.14	35	<0.6
LAW-61	30.9	0.8	<0.5	11	52	-	-	<0.06	0.98	<0.6
LAW-62	30.4	<0.6	<0.6	7.9	39	-	-	<0.07	1.4	<0.7
LAW-63	26.9	<0.6	<0.6	16	48	-	-	0.5	44	<0.7

Table 4. (continued)

Reference No.	% Dry Matter	Cr	Ni	Cu	Zn	As	Se	Cd	Hg	Pb
P. vitulina										
LAW-64	25.8	<0.5	<0.5	8.9	51	-	-	0.14	12	<0.6
LAW-65	28.7	0.6	<0.5	17	79	-	-	2.9	95	<0.6
LAW-66	28.3	<0.5	<0.5	11	39	-	-	<0.06	0.99	<0.6
LAW 67	30.8	<0.5	<0.5	11	29	-	-	0.13	5.7	<0.6
LAW 68	24.6	<0.5	<0.5	26	60	-	-	<0.06	1	<0.6
LAW 69	23.1	<0.5	<0.5	7.2	25	-	-	0.16	5	0.18
B. acutorostrata										
SW1991/2	23.7	2.1	1.2	5	35	-	-	1.1	1.8	0.15
D. delphis										
IC1990/4	29.6	<0.56	<0.56	5.8	35	-	8.2	0.24	0.57	0.12
IC1991/1	24.1	1.7	1.2	4	45	-	-	0.33	66	0.14
IC1991/1	17.6	<0.5	<0.5	40	15	-	-	<0.06	0.5	0.05
SW1990/5A	27.6	<0.5	<0.5	7.1	42	-	-	0.71	6.1	0.43
SW1990/26	25.9	<0.5	<0.7	4.2	26	-	-	0.34	11	1
SW1990/45	27.5	<0.5	<0.5	12	89	-	-	2.2	6.9	0.37
SW1990/46	30.3	<0.55	<0.55	3.9	58	-	2.7	1.1	0.14	0.19
SW1990/47	25.7	<0.65	<0.65	9.6	67	-	13	9	1.9	0.29
SW1990/68	29.3	0.92	<0.57	3.1	41	-	2.3	0.18	11	0.09
SW1990/78	28.2	<0.51	<0.51	5.1	42	-	13	0.68	67	0.11
SW1990/81	27	<0.5	<0.5	5.3	45	-	na	0.1	3.7	0.21
SW1990/87	27.5	<0.57	<0.56	8.4	36	-	2.9	0.21	5	0.13
SW1990/88	25	<0.57	<0.53	81	19	-	1.7	<0.06	3.4	0.1
SW1990/90A	27.3	<0.51	<0.51	6.9	36	-	6.2	0.54	13	0.04
SW1990/97	31.6	<0.52	<0.52	3.6	46	-	1.3	<0.07	0.21	0.05
SW1990/101	27.2	<0.52	<0.52	7	30	-	4.7	0.32	8.3	0.09
SW1990/102	25.3	<0.52	<0.52	4.3	32	-	1.2	0.29	2.1	0.08
SW1990/103	27.9	<0.57	<0.57	4.9	30	-	2.1	0.33	1.7	0.07
SW1990/104	26.7	<0.51	<0.51	4.3	30	-	3.1	0.31	3.6	0.07
SW1990/108	25.9	<0.58	<0.58	4.4	38	-	13	1	30	0.08
SW1991/7	28.1	<0.68	<0.62	6.8	32	-	38	0.6	93	0.13
SW1991/10	28.3	<0.5	<0.5	3.2	32	-	-	0.09	2.5	0.28
SW1991/37	28.2	<0.5	<0.5	4.4	35	-	-	0.34	11	0.19
SW1991/40	27.7	<0.51	<0.51	4.9	28	-	11	0.34	25	0.07
SW1991/46	24.7	0.76	<0.54	4.5	37	-	22	0.12	2.2	0.08
SW1991/83	29.1	<0.55	<0.55	4.2	25	-	45	0.3	130	0.1
SW1991/129	31.2	<0.57	<0.57	6.4	99	-	0.8	<0.07	0.67	0.06
SW1991/137	28.2	1.1	<0.5	5.1	43	-	12	6.5	33	0.03
SW1992/1	22.1	<0.56	<0.56	2.7	21	0.86	-	2.7	3.1	0.12
SW1992/2	26.2	0.74	<0.67	4.9	32	0.7	-	0.16	11	0.05
SW1992/3	25.1	<0.68	<0.68	5.3	37	0.91	-	0.12	7.4	0.05
SW1992/4	30.5	<0.57	<0.57	4.2	24	0.73	-	0.14	6.1	0.04
SW1992/10	27.9	<0.65	<0.65	5.6	32	0.89	3.8	0.72	6.4	0.12
SW1992/11	27.6	<0.57	<0.5	4.1	64	2.6	2.2	0.48	2.8	0.02
SW1992/12	26.7	<0.52	<0.52	5.1	30	0.51	8.5	0.94	24	0.05
SW1992/14(1)	29.2	<0.63	<0.63	4.6	34	1.6	2.5	0.6	2.7	0.14
SW1992/14(2)	24.9	<0.61	<0.61	6	36	1.5	4.1	0.65	8.1	0.03
SW1992/19(1)	26.6	0.64	<0.62	5.1	34	1.4	-	0.47	0.3	0.06
SW1992/19(2)	25.3	<0.62	<0.62	5.7	27	0.85	-	0.47	5.6	0.09
SW1992/20	26.7	<0.54	<0.54	4.7	31	0.77	-	0.72	11	0.15
SW1992/172(1)	30.7	0.81	<0.62	6.5	24	-	18	0.66	50	0.11
SW1992/172(2)	34.5	1	<0.57	5.3	37	-	1.7	<0.07	1.5	0.1
G. griseus										
SW1990/70	23.5	0.7	0.6	31	40	-	-	<0.06	1.3	0.15
G. melas										
SW1990/88A	30.4	<0.65	<0.65	9.8	41	-	6.9	0.34	25	0.1
SW1991/55	28.2	5.6	1.1	3.8	65	-	39	65	23	0.22
L. acutus										
LAW 70	36.1	<0.5	<0.5	6.7	36	-	-	1.8	44	<0.6
SW1990/114	26.2	<0.54	<0.54	6.2	71	-	1.3	<0.07	0.39	<0.03
SW1991/9	26.3	2.8	<0.57	8.1	61	-	33	0.76	72	0.08

Table 4. (continued)

Reference No.	% Dry Matter	Cr	Ni	Cu	Zn	As	Se	Cd	Hg	Pb
L. albirostris										
SW1989/49	30	<0.6	<0.6	6.8	27	-	-	0.11	27	3.8
SW1991/108	30.4	0.56	<0.54	6.7	34	-	2.1	0.08	0.8	0.11
SW1991/126	27.9	<0.55	<0.55	5.1	26	-	4.9	0.07	4.1	0.13
S. coeruleoalba										
SW1990/4	26.6	<0.5	<0.6	10	65	-	-	8.4	10	<0.7
SW1990/20	26.8	<0.5	<0.5	12	46	-	-	11	11	<0.6
SW1990/57	27.9	0.69	0.73	3.7	8.5	-	0.86	<0.07	0.95	<0.03
SW1991/11	32.5	3.7	<0.53	7.4	42	-	9.1	4.6	22	0.06
SW1991/18	30.1	0.66	<0.51	6.4	51	-	4.2	4.8	4.4	0.11
SW1991/26	27.7	<0.5	<0.5	8.4	45	-	-	1.7	5.7	1.6
SW1991/122	29.2	<0.59	<0.59	5.1	39	-	0.94	0.21	0.59	0.1
T. truncatus										
LAW-71	23.3	<0.6	<0.6	5.7	32	-	-	0.07	20	<0.7
SW1989/47	26.3	<0.5	<0.5	8.3	42	-	-	0.12	21	<0.6
SW1991/42	27.1	0.68	<0.63	5.2	29	-	83	<0.08	201	0.58
SW1991/115	28.2	<0.55	<0.55	13	268	-	0.97	<0.07	1.6	0.07
SW1991/130	36.3	<0.59	<0.59	3.8	47	-	40	<0.07	93	0.14
SW1992/25	33.4	<0.52	<0.52	8.1	37	-	81	0.09	191	0.61
P. phocoena										
IC1990/1	32	0.26	0.05	5.7	54	-	2.6	0.04	1.8	0.2
IC1990/2	30.7	<0.5	1	11	40	-	-	0.08	22	0.37
IC1990/3	29.8	<0.5	0.5	5	40	-	-	<0.06	0.6	0.26
IC1990/4	28.8	<0.5	<0.5	27	46	-	-	<0.06	0.7	0.21
91L-2947	36.1	<0.52	<0.52	53	28	-	1.3	<0.06	<0.01	0.06
SW1989/43A	30	<0.6	<0.6	13	140	-	-	0.23	150	4.3
SW1989/51	22.9	0.84	<0.54	46	57	-	2.9	<0.06	0.07	<0.03
SW1989/89	27.9	<0.4	<0.6	12	50	-	-	0.41	10	<0.7
SMRU90-04	29.2	0.26	<0.02	21	37	-	1.2	0.03	0.3	0.04
SMRU90-51	27	0.82	<0.49	8.8	29	-	3.3	0.11	3.8	0.39
SMRU90-55	34	1.2	0.76	4.2	115	-	6	<0.06	12	0.07
SMRU90-60	31.6	0.79	<0.53	5.4	22	-	2	0.07	0.26	<0.03
SMRU90-61	30.4	0.69	<0.6	6.1	37	-	3.9	0.16	10	<0.03
SMRU90-72A	26.7	0.45	0.12	12	84	-	54	0.66	102	0.26
SW1990/31	28.9	<0.6	<0.5	11	90	-	-	0.18	16	<0.6
SW1990/39	22.9	1.6	0.7	4.1	26	-	-	<0.06	0.5	0.52
SW1990/50	27.5	<0.5	<0.5	12	32	-	-	0.22	14	0.25
SW1990/50A	27.8	<0.5	<0.5	6.5	23	-	-	<0.06	0.8	0.17
SW1990/51(1)	35.8	<0.5	<0.5	8.7	39	-	-	0.34	57	0.46
SW1990/51(2)	27.5	<0.5	<0.5	54	80	-	-	<0.06	2	0.18
SW1990/54	33.9	<0.5	<0.5	26	39	-	-	<0.06	1.8	0.19
SW1990/65	26.1	<0.5	<0.5	11	150	-	-	0.21	30	0.29
SW1990/70	23.5	0.7	0.6	31	40	-	-	<0.06	1.3	0.15
SW1990/71	26.8	0.77	<0.53	8.2	34	-	6.6	0.13	10	0.16
SW1990/72	26.3	0.9	0.9	7.8	42	-	-	0.22	12	0.16
SW1990/72A	23.5	0.7	0.6	31	40	-	-	<0.06	1.3	0.15
SW1990/77A	27.7	0.9	<0.5	8.4	30	-	-	0.44	15	0.12
SW1990/77B	26.4	<0.5	<0.5	7.5	31	-	-	0.37	20	0.11
SW1990/79	24.6	<0.52	<0.52	7.6	35	-	1.3	0.13	0.94	<0.03
SW1990/84	30.1	<0.5	<0.5	6	32	-	1.1	0.93	0.57	<0.03
SW1990/92	25.5	<0.5	1	7.6	120	-	-	0.14	73	0.49
SW1990/94	25.7	<0.62	<0.55	7.6	73	-	12	0.16	45	0.03
SW1990/96	27.7	<0.54	<0.54	6.9	60	-	5	0.27	11	0.13
SW1990/98	27.1	<0.5	<0.5	5.7	110	-	-	0.18	4.3	0.2
SW1990/100	25.1	<0.5	1.1	12	110	-	-	0.07	23	0.2
SW1990/100A	27.7	0.76	<0.53	8.2	43	-	0.8	<0.06	0.65	<0.03
SW1990/106	24.5	<0.53	<0.53	8.4	104	-	-	0.1	12	0.07
SW1990/107	23.8	1.8	0.9	17	110	-	-	0.44	190	0.16
SW1990/109	35.1	<0.52	<0.52	4	26	-	0.9	0.2	1.1	<0.03
SW1990/111	30.2	<0.5	0.7	5.9	30	-	-	0.11	6.8	0.18
SMRU91-06	28.1	0.53	<0.5	6.3	55	-	12	0.23	28	0.07

Table 4. (continued)

Reference No.	% Dry Matter	Cr	Ni	Cu	Zn	As	Se	Cd	Hg	Pb
P.phocoena										
SMRU91-15	29.6	0.64	0.17	13	66	-	1.1	<0.02	0.71	0.07
SMRU91-16	25.2	0.67	0.25	36	162	-	1.6	0.02	1.6	0.06
91L-2937	40	<0.53	<0.53	11	42	-	2	<0.06	1.4	0.05
91L-2936	31.3	<0.51	<0.51	7.7	37	-	5.6	0.12	2.3	0.56
SMRU91-07	29.6	<0.54	<0.54	4.8	25	-	2.6	0.23	3	<0.03
SW1991/14	24.7	<0.5	<0.5	4.5	174	-	-	0.25	52	0.06
SW1991/17	25.8	4.4	<0.54	8.5	104	-	4.8	0.43	8.6	0.03
SW1991/17A	23.5	0.69	<0.55	4.3	41	-	3.9	0.15	7	0.05
SW1991/19A	25.6	<0.51	<0.51	13	53	-	8.1	1.4	24	<0.03
SW1991/19B	26.2	0.8	<0.57	6.9	30	-	1.1	<0.07	0.12	0.08
SW1991/20	25.1	<0.52	<0.52	9.8	181	-	2.6	0.71	4.1	<0.03
SW1991/22	25.7	0.72	<0.51	7.8	76	-	7.6	0.51	31	<0.03
SW1991/23	27.2	0.48	0.14	6.2	70	-	1.7	0.03	0.89	0.03
SW1991/24	26.2	<0.5	<0.5	5.3	31	-	5.1	0.6	12	<0.03
SW1991/28	27.2	<0.52	<0.52	6.6	29	-	2	0.31	2.1	0.05
SW1991/29	27.5	0.77	0.68	14	40	-	1	0.27	0.54	0.1
SW1991/30	29.8	0.24	<0.02	3.8	46	-	1.1	0.21	0.58	0.05
SW1991/31	27.2	0.27	0.04	5.4	41	-	2.7	0.54	3	0.07
SW1991/32A	26.9	0.98	<0.6	7.2	33	-	2.4	<0.07	4.3	0.04
SW1991/35	26.2	<0.57	<0.57	15	110	-	1.7	<0.07	1.5	0.07
SW1991/36	25.9	<0.52	<0.52	15	27	-	47	0.55	82	0.16
SW1991/43	26.9	0.68	<0.53	9.5	33	-	5.3	0.26	8.5	0.06
SW1991/48	24.5	0.69	<0.49	9	50	-	4.3	0.18	0.74	0.1
SW1991/54	28	<0.58	<0.58	10	63	-	8.4	<0.07	14	0.14
SW1991/59	24.5	<0.56	<0.56	76	50	-	2.3	<0.07	1.8	0.05
SW1991/61	21.1	0.8	<0.5	2.8	20	-	-	<0.06	1	0.08
SW1991/63	41.6	0.91	0.26	68	36	-	4.4	<0.02	1.1	0.04
SW1991/70	28.7	0.65	0.15	7.6	65	-	3	0.03	2.3	0.09
SW1991/71	26.8	1.2	<0.5	8.8	86	-	-	0.09	5.9	0.35
SW1991/73	26.3	<0.56	<0.56	24	41	-	0.94	<0.07	1.5	<0.03
SW1991/74	27.2	<0.53	<0.53	66	77	-	0.86	<0.06	1.2	0.04
SW1991/84	48.1	<0.55	<0.55	70	38	-	1.9	<0.07	1.2	0.06
SW1991/95	30.6	1.1	<0.54	12	34	-	17	0.29	28	0.22
SW1991/96	29.8	0.47	0.07	13	36	-	1.7	<0.02	0.64	0.02
SW1991/104	23.9	<0.5	<0.5	11	77	-	7	0.26	24	<0.03
SW1991/109	26.6	<0.59	<0.59	8.4	44	-	1.8	0.19	1.6	0.07
SW1991/111	28.2	<0.57	<0.57	6.9	33	-	38	2.8	99	0.09
SW1991/116	23.7	<0.5	<0.5	3.3	20	-	1.7	0.68	1.3	<0.03
SW1991/117A	30.6	0.38	0.09	9.2	5.5	-	2.1	0.28	1	0.06
SW1991/120	25	0.74	<0.54	7.4	137	-	1.8	<0.08	3.5	0.06
SW1991/124	25.3	<0.52	<0.52	6.9	51	-	2.9	<0.06	3.4	0.04
SW1991/128	25.4	0.3	0.04	7.5	57	-	2.2	0.17	2.5	0.03
SW1991/135	27.1	<0.53	<0.53	57	40	-	1.2	<0.06	0.74	0.18
SW1991/136	28.9	<0.54	<0.54	13	43	-	106	0.25	65	0.63
SW1991/142	22.9	0.52	<0.49	3.5	20	-	1.1	0.08	<0.01	<0.03
SW1992/7	28.4	<0.51	<0.51	9.6	35	-	3.7	<0.06	2.5	0.08
SW1992/112	20.6	<0.5	<0.5	2.9	17	-	2.8	0.15	0.83	0.04
SW1992/170	22.2	<0.53	<0.53	5	23	-	17	0.84	1.2	0.09
SW1992/175	30.7	1.9	<0.58	10	28	-	17	<0.07	31	0.12
LAW-01	27.5	<0.6	<0.6	12	50	-	-	0.07	1.4	<0.7
LAW-02	26.4	<0.6	<0.6	8.9	48	-	-	0.16	8.4	<0.7
LAW-03	28.7	<0.6	<0.6	160	49	-	-	<0.07	0.66	<0.7
LAW-04	31.8	<0.6	<0.6	120	62	-	-	<0.07	2.9	<0.7
LAW-05	40.5	<0.6	<0.6	8.7	49	-	-	<0.07	0.63	<0.7
LAW-06	55.5	<0.5	<0.5	6.6	25	-	-	<0.06	0.61	<0.6
LAW-07	27.2	<0.6	<0.6	17	66	-	-	0.11	0.59	<0.7
LAW-08	27.4	<0.6	<0.6	6.5	44	-	-	0.24	26	<0.7
LAW-09	27.1	<0.6	<0.6	16	120	-	-	0.12	5.2	<0.7
LAW-10	28.6	<0.6	<0.6	8.1	140	-	-	0.14	30	<0.7
LAW-11	30.2	<0.6	<0.6	10	36	-	-	0.11	7.5	<0.7
LAW-12	31.1	<0.6	<0.6	8.2	31	-	-	0.14	5.4	<0.7
LAW-13	28.3	<0.6	<0.6	10	34	-	-	0.17	4.9	<0.7
LAW-14	30.3	<0.6	<0.6	7.4	34	-	-	1.2	1.1	<0.7
LAW-15	25	<0.5	<0.5	7.3	87	-	-	<0.06	2.8	<0.6
LAW-16	30.2	0.5	<0.5	30	71	-	-	<0.06	1	<0.6
LAW-17	28.4	<0.6	<0.6	17	36	-	-	<0.07	0.89	<0.7
LAW-18	21.6	<0.5	<0.5	79	40	-	-	<0.06	1.6	0.11

Table 5. Concentrations of trace metals in kidney samples (mg kg⁻¹ wet weight)

ReferenceNo.	% Dry Matter	Cr	Ni	Cu	Zn	As	Se	Cd	Hg	Pb
H. grypus										
SS1991/28	21.3	<0.55	<0.55	5.2	40	-	8.8	0.43	22	0.31
SS1991/30A	27.9	<0.5	<0.5	12	28	-	31	<0.06	13	0.86
SS1991/55	22.6	<0.57	<0.57	5.7	47	-	0.29	<0.07	<0.01	<0.03
SS1991/58	24.3	<0.55	<0.55	2.7	48	-	2.6	0.71	5.2	<0.03
SS1991/73	20.4	<0.49	<0.49	8.7	51	-	0.52	<0.06	2.9	0.05
SS1992/9	22.8	<0.57	<0.57	2.0	27	-	1.4	0.7	0.88	0.04
D.delphis										
SW1991/40	31.9	<0.56	<0.56	2.1	15	-	2.6	1.6	2.3	0.03
SW1991/46	20.9	0.63	<0.58	3.1	20	-	3.7	1.9	0.48	0.05
SW1991/83	20.9	<0.58	<0.58	2.6	17	-	3.3	4.1	4.4	0.08
SW1991/129	18.8	<0.54	<0.54	7.6	21	-	2.3	<0.06	0.35	<0.03
SW1992/1	21.5	<0.58	<0.58	3.0	19	0.87	-	2.9	3.5	0.11
SW1992/2	23.7	1.1	0.58	2.5	16	0.83	-	1.1	1.5	0.02
SW1992/3	25.3	<0.66	<0.66	5.4	35	0.84	-	0.12	6.0	0.04
SW1992/4	27.4	0.83	<0.67	2.7	17	1.4	-	1.4	1.2	0.03
SW1992/10	27.1	2.2	<0.55	2.5	15	-	3.9	2.3	0.86	0.03
SW1992/11	21.3	<0.52	<0.52	3.4	22	-	2.1	3.0	1.1	<0.03
SW1992/12	26.7	<0.56	<0.56	2.5	19	-	3.6	9.3	2.7	<0.03
SW1992/14(1)	26.1	<0.57	<0.57	3.1	19	-	3.0	2.0	0.72	<0.03
SW1992/14(2)	23.9	<0.54	<0.54	2.5	15	-	1.9	1.4	0.8	<0.03
SW1992/19(1)	22.8	2.6	1.3	2.7	16	0.95	-	2.6	0.3	0.05
SW1992/19(2)	24.1	<0.57	<0.57	3.1	17	1.0	-	2.7	1.5	0.06
SW1992/20	28.5	3.1	1.6	1.9	12	1.4	-	2.4	0.9	0.06
SW1992/172(1)	21.4	1.0	<0.5	3.0	17	-	3.6	3.3	4.7	0.07
SW1992/172(2)	22.0	2.2	<0.67	2.9	17	-	2.2	0.26	0.54	0.06
G.melas										
SW1991/55	20.8	1.2	<0.56	8.5	51	-	11	176	12	0.06
L. albirostris										
SW1991/108	23.0	<0.59	<0.59	3.1	19	-	2.9	0.67	0.11	0.04
SW1991/126	23.4	<0.57	<0.57	3.4	15	-	2.6	0.36	1.3	<0.03
S.coeruleoalba										
SW1991/122	23.9	<0.62	<0.62	4.2	24	-	2.3	1.0	0.42	0.09
T. truncatus										
SW1991/42	26.7	0.77	<0.59	2.0	17	-	9.9	0.2	18	0.17
SW1991/115	22.1	<0.52	<0.52	9.3	66	-	0.91	<0.06	0.85	0.05
SW1991/130	32.7	<0.51	<0.51	1.7	20	-	4.7	0.15	6.0	0.05
SW1992/25	25.0	<0.55	<0.55	2.6	22	-	6.8	1.2	11	0.19
P.phocoena										
SW1991/19B	23.3	<0.53	<0.53	3.0	16	-	2.3	<0.06	1.1	0.03
SW1991/32A	23.9	1.1	<0.59	2.3	16	-	1.9	0.26	1.2	0.06
SW1991/35	22.7	<0.53	<0.53	3.3	22	-	2.3	0.24	<0.01	<0.03
SW1991/36	20.0	<0.57	<0.57	2.9	17	-	2.4	2.5	4.9	0.03
SW1991/43	18.8	1.8	<0.68	3.6	18	-	2.5	1.7	0.81	0.06
SW1991/48	20.7	0.74	<0.56	4.6	21	-	1.5	1.2	0.19	0.06
SW1991/54	20.6	<0.55	<0.55	3.9	20	-	2.6	0.7	1.5	0.08
SW1991/59	18.8	<0.52	<0.52	3.1	19	-	0.58	<0.06	0.59	<0.03
SW1991/73	21.3	<0.54	<0.54	5.9	18	-	0.64	<0.06	0.46	<0.03
SW1991/74	20.3	<0.56	<0.56	4.7	21	-	0.58	<0.07	0.5	0.07
SW1991/95	25.6	0.85	<0.6	3.5	21	-	2.7	0.9	3.7	0.07
SW1991/104	27.0	<0.53	<0.53	9.0	33	-	12	0.18	32	0.03
SW1991/109	24.3	<0.53	<0.53	4.6	22	-	3.2	0.76	0.49	0.03
SW1991/111	24.2	<0.51	<0.51	3.5	21	-	4.9	8.7	9.5	0.04
SW1991/116	20.8	<0.58	<0.58	3.7	23	-	2.4	0.64	4.3	0.05
SW1991/120	19.2	<0.51	<0.55	3.2	22	-	0.86	0.38	0.37	0.03
SW1991/135	20.6	<0.55	<0.55	3.5	19	-	0.86	0.07	<0.01	<0.03
SW1991/136	20.1	<0.53	<0.53	3.1	22	-	2.2	1.5	5.3	0.11
SW1991/142	23.8	<0.55	<0.55	12	29	-	0.8	<0.07	<0.01	<0.03
SW1992/7	19.8	1.2	<0.51	2.9	19	-	1.6	0.12	0.81	<0.03
SW1992/112	27.9	0.63	<0.54	15	29	-	3.0	0.3	37	0.09
SW1992/170	28.3	<0.49	<0.49	13	45	-	1.3	0.18	5.6	0.23
SW1992/175	22.0	2.0	<0.59	3.3	18	-	2.6	0.87	2.1	0.09

Table 6. Concentrations of trace metals in stomach contents of common dolphins (mg kg⁻¹ wet weight)

ReferenceNo.	Cr	Ni	Cu	Zn	As	Se	Cd	Hg	Pb
D.delphis									
SW1992/2	<0.87	<0.87	1.7	5.9	1.3	-	0.77	0.2	<1.1
SW1992/4	6.1	2.9	1.5	14	2	-	0.75	0.24	<1.0
SW1992/19(1)	<0.62	0.78	1.7	61	3.5	-	0.6	0.1	<0.75
SW1992/19(2)	<0.71	<0.71	4.1	103	0.62	-	0.65	0.09	<0.85
SW1992/20	13	6.4	2.9	46	1.1	-	0.84	0.09	<1.2

Table 7. Concentrations of total chlorobiphenyls and organochlorines other than chlorobiphenyl congeners in blubber samples (mg kg⁻¹ wet weight)

Reference No.	% HEL	ICES 7 CBs	Total 25 CBs	HCB	α-HCH	γ-HCH	p,p'-DDE	p,p'-DDT	p,p'-TDE	Dieldrin
H. grypus										
SS1991/23	41	38.6	55.7	-	-	-	4.8	-	-	-
D. delphis										
SMRU90-73	91	5.31	9.34	0.56	0.002	nd	8.3	0.05	0.036	0.027
IC1990/4	85	26.9	47.3	0.18	0.058	0.24	7.1	0.26	1.1	1.9
IC1991/1	75	1.35	2.34	0.012	-	-	0.8	-	-	-
SW1990/5A	71	15	24.9	0.23	-	-	5.7	-	-	-
SW1990/20A	55	18.8	33.2	0.18	0.053	0.15	4.3	0.19	0.63	1.2
SW1990/45	75	6.09	9.53	0.34	-	-	7.3	-	-	-
SW1990/46	90	8.42	15.5	0.58	-	-	5.9	-	-	-
SW1990/47	82	3.46	10.9	0.11	-	-	1.2	-	-	-
SW1990/68	85	49	83.8	0.37	-	-	34.4	-	-	-
SW1990/78	86	46.6	82.1	0.26	0.045	0.13	24.9	0.14	2	1.9
SW1990/81	82	1.41	2.26	0.014	-	-	0.51	-	-	-
SW1990/87	88	14.5	26.1	0.13	-	-	3.3	-	-	-
SW1990/88	78	10.2	16.1	0.3	0.039	0.068	5.5	0.11	0.55	1.4
SW1990/97	79	43.4	75.8	0.64	0.12	0.59	17.2	0.54	1.7	5.3
SW1990/101	82	19.8	34.1	0.16	0.056	0.16	6.2	0.25	1.2	1.7
SW1990/102	82	18.5	31.8	0.13	0.039	0.11	5.4	0.2	0.71	1.2
SW1990/103	80	29.2	50	0.16	0.038	0.11	13.9	0.23	1	1.7
SW1990/104	80	20.3	35.6	0.19	-	-	5.2	-	-	-
SW1990/108	91	20.7	35.7	0.19	0.047	0.14	11	0.22	0.14	1.7
SW1991/7	87	3.06	5.47	0.025	0.044	0.13	0.27	0.14	0.81	1.5
SW1991/10	86	36	57.2	0.42	-	-	29	-	-	-
SW1991/37	78	19.2	35.2	0.28	0.089	0.28	3.8	1.2	1.9	2.9
SW1991/83	90	4.7	8.26	0.036	-	-	0.49	-	-	-
SW1991/137	76	12.7	23.6	0.12	0.032	0.26	3.9	0.44	0.8	0.85
SW1992/1	82	41.6	71.4	0.068	0.26	0.19	2.4	11	2.2	1.8
SW1992/2	80	19.8	34	0.087	0.24	0.19	2.5	5.3	1.4	0.78
SW1992/3	84	27.8	47.8	0.073	0.18	0.19	2.6	7.5	1.7	1
SW1992/4	84	21.1	35.7	0.08	0.21	0.2	1.9	5	1.1	0.77
SW1992/10	92	22.4	38.1	0.058	0.16	0.16	1.4	5.4	1.1	1
SW1992/11	88	11.1	18.9	0.06	0.22	0.21	1.2	2.1	0.6	0.54
SW1992/12	64	18.7	31	nd	0.064	0.11	0.71	8.3	0.57	0.44
SW1992/14(1)	90	11.3	19.3	0.048	0.092	0.13	0.72	2.8	0.49	0.45
SW1992/14(2)	92	8.35	14.2	0.06	0.11	0.15	0.87	2.7	0.57	0.6
SW1992/19(1)	84	8.88	15.2	0.079	0.25	0.27	1.3	1.7	0.6	0.5
SW1992/19(2)	86	11.9	20	0.06	0.1	0.12	1.1	5.9	1	1
G. griseus										
SW1990/70	88	6.49	10.1	0.12	0.015	0.047	1.2	0.34	0.99	0.94
G. melas										
SW1990/88A	84	29.1	44.8	0.13	-	-	11.3	-	-	-
SW1991/55	75	17.8	28.9	1.02	-	-	16.6	-	-	-
L. acutus										
SW1989/88B	77	13.5	27	-	0.22	0.11	-	4.2	4.3	4
S. coerulealba										
SW1991/26	56	10.1	18.4	0.55	-	-	8.2	-	-	-
T. truncatus										
SW1991/42	68	72.1	119.3	0.22	-	-	36.8	-	-	-
SW1991/115	57	6.87	10.9	0.36	-	-	3.3	-	-	-
P. phocoena										
IC1990/1	84	4.88	9.36	0.37	-	-	1.5	-	-	-
IC1990/2	73	0.92	1.44	0.037	-	-	0.29	-	-	-
IC1990/3	70	10.4	16.1	0.43	-	-	2.5	-	-	-
IC1990/4	93	5.17	7.93	0.27	-	-	1.1	-	-	-
IC1991/2	62	4.46	7.33	0.26	nd	0.026	1.5	0.26	0.42	nd
SMRU90-01	86	6.68	12.2	0.26	-	-	3.3	-	-	-
SMRU90-04	93	2.16	3.64	0.19	-	-	1	-	-	-
SMRU90-02	89	0.17	0.26	0.011	-	-	0.11	-	-	-
SMRU90-03	82	5.57	9.62	0.29	-	-	2.8	-	-	-
SMRU90-05	87	4.13	5.92	0.31	-	-	3.2	-	-	-
SMRU90-06	89	3.61	5.24	0.36	-	-	2.4	-	-	-
SMRU90-07	83	0.11	0.17	0.002	-	-	0.064	-	-	-
SMRU90-08	91	4.15	6.02	0.39	-	-	2.8	-	-	-
SMRU90-09	87	4.58	6.82	0.46	-	-	2.9	-	-	-
SMRU90-10	72	2.34	3.86	0.25	-	-	1.4	-	-	-
SMRU90-11	90	0.086	0.12	0.009	-	-	0.057	-	-	-
SMRU90-12	87	0.65	0.65	nd	-	-	0.39	-	-	-
SMRU90-13	76	3.38	4.9	0.31	-	-	1.9	-	-	-
SMRU90-14	89	5.62	8.29	0.63	-	-	4.7	-	-	-
SMRU90-15	92	5.51	8.23	0.27	-	-	4.3	-	-	-

Table 7. (continued)

ReferenceNo.	% HEL	ICES 7 CBs	Total 25 CBs	HCB	α -HCH	γ -HCH	p,p'-DDE	p,p'-DDT	p,p'-TDE	Dieldrin
P. phocoena										
SMRU90-16	91	0.18	0.26	0.012	-	-	0.16	-	-	-
SMRU90-17	87	8.45	13	0.39	-	-	5.1	-	-	-
SMRU90-18	93	9.93	15.2	0.44	-	-	5.6	-	-	-
SMRU90-19	89	6.12	9.08	0.43	-	-	4.4	-	-	-
SMRU90-51	93	12.9	23.1	0.092	0.018	0.21	2.3	1.8	0.88	1
SMRU90-55	85	43.3	76.9	0.94	0.07	0.31	17	5.7	5.4	11.4
SMRU90-60	92	0.085	0.12	0.005	-	-	0.04	-	-	-
SMRU90-61	89	10.6	17.1	1.8	-	-	5.5	-	-	-
SMRU90-75	93	6.96	12.9	0.61	0.13	0.21	3.7	2	1.8	3.3
SMRU90-76	89	9.68	15.4	0.54	-	-	6.5	-	-	-
SW1990/39	92	11	18.6	0.35	-	-	2.1	-	-	-
SW1990/50	95	9.99	15.9	0.23	-	-	1.3	-	-	-
SW1990/50A	79	6.87	9.96	0.44	-	-	1.6	-	-	-
SW1990/51(1)	78	3.07	5.88	0.026	0.006	0.022	0.28	0.092	0.13	0.22
SW1990/51(2)	70	0.2	0.34	0.031	0.004	0.02	0.22	0.011	0.11	0.2
SW1990/54	64	12.5	22.3	0.38	0.079	0.17	2.9	1	1.6	1.4
SW1990/65	76	23.2	36.4	0.98	-	-	6.3	-	-	-
SW1990/66	85	16	25.5	0.45	-	-	2.6	-	-	-
SW1990/71	88	2.06	3.39	0.025	0.006	0.02	0.92	0.83	0.29	0.29
SW1990/72	81	19	32.9	0.3	0.016	0.11	2.9	1.4	2.5	3.6
SW1990/72A	88	6.49	10.1	0.12	0.015	0.047	1.2	0.34	0.99	0.94
SW1990/73	71	18.1	31.3	0.5	-	-	2.6	-	-	-
SW1990/77B	85	27.9	52.9	0.37	-	-	2.6	-	-	-
SW1990/79	87	2.84	4.69	0.32	0.027	0.012	1.6	0.65	0.57	1.6
SW1990/84	91	17.9	29.9	0.3	0.02	0.077	7.9	7.1	5.8	8.6
SW1990/92	83	0.19	0.33	0.005	-	-	0.076	-	-	-
SW1990/94	75	13.3	24.4	0.32	0.039	0.11	6.6	4	2.8	6.6
SW1990/96	85	43.7	76.9	0.73	0.077	0.31	8.9	4.5	3.8	7.7
SW1990/98	85	1.11	1.71	0.049	-	-	0.39	-	-	-
SW1990/100	76	0.8	1.24	0.037	-	-	0.25	-	-	-
SW1990/100A	89	8	13.3	1.1	0.15	0.23	7	3.6	2.7	8
SW1990/106	76	21.2	34.8	0.53	0.066	0.28	7.1	0.1	2.1	4
SW1990/107	87	4.79	8.05	0.068	-	-	0.93	-	-	-
SW1990/109	89	14.2	24.7	0.2	0.078	0.17	2.1	0.14	0.79	0.93
SW1990/111	93	0.24	0.41	0.003	-	-	0.049	-	-	-
SMRU91-06	92	2.38	4.09	0.13	0.067	0.1	1.1	0.11	1	3
SMRU91-08	90	1.14	1.95	0.063	nd	nd	0.44	0.59	0.21	0.15
SMRU91-15	92	13.5	21.8	0.64	0.11	0.35	3.8	1.2	3.5	5
SMRU91-07	93	2.64	4.63	0.21	nd	nd	1.2	nd	0.75	nd
SW1991/14	64	52.7	88.8	0.54	0.066	0.16	16.8	0.25	4.7	5.6
SW1991/17	89	24.2	41.1	0.44	0.061	0.16	10.5	0.12	3	6.3
SW1991/17A	87	18.1	32.8	0.38	0.058	0.23	4.1	0.085	2.4	5
SW1991/19A	90	18.2	31.7	0.4	0.06	0.17	6.3	0.12	2.6	5.6
SW1991/19B	89	8.13	14.3	0.2	0.074	0.18	1.7	0.77	1.1	0.97
SW1991/20	87	7.11	12.9	0.45	0.074	0.22	2.9	0.12	0.98	2.5
SW1991/22	85	11.3	19	0.29	nd	0.055	5.2	0.032	0.87	0.16
SW1991/23	86	30	50.8	0.83	0.14	0.44	8.1	1.9	3.2	6.6
SW1991/24	91	21.4	37.8	0.5	0.078	0.3	7.3	0.14	1.3	6
SW1991/28	89	3.96	7.06	0.46	0.045	0.058	2.6	0.038	0.62	2
SW1991/29	88	6.22	10.5	0.49	0.037	0.057	3.7	0.11	0.09	3.3
SW1991/30	92	4.19	7.19	0.28	0.11	0.23	1.6	0.58	0.63	1.5
SW1991/31	87	22.8	38.4	0.49	0.088	0.24	8.4	2	2.7	4.3
SW1991/32A	91	6.37	11	0.14	0.035	0.081	1	0.48	0.45	0.62
SW1991/35	74	4.91	7.46	0.39	-	-	1.5	-	-	-
SW1991/36	93	6.7	12	0.22	0.048	0.2	1.9	0.11	0.93	1.8
SW1991/43	88	2.05	4.19	0.2	0.045	0.052	1.1	0.58	0.46	0.68
SW1991/48	86	5.48	8.5	0.14	-	-	1.2	-	-	-
SW1991/54	90	11	17.7	0.33	-	-	3.1	-	-	-
SW1991/59	77	3.25	5.9	0.13	-	-	0.77	-	-	-
SW1991/61	52	1.78	3.05	0.086	-	-	0.25	-	-	-
SW1991/63	75	4.81	8.56	0.29	0.034	0.15	2.2	0.86	1.6	4.3
SW1991/70	92	16.6	25.3	0.43	0.094	0.45	2.2	0.67	1.7	2.4
SW1991/71	81	11.3	19.4	0.58	-	-	2.7	-	-	-
SW1991/73	53	5.15	9.06	0.2	-	-	1.8	-	-	-
SW1991/74	77	2.87	5.05	0.076	-	-	0.28	-	-	-
SW1991/95	84	10.1	17.1	0.29	-	-	3.2	-	-	-
SW1991/100	62	4.39	7.75	0.22	0.031	0.14	1.7	0.41	2.9	3.5
SW1991/104	86	32.5	58.1	0.8	-	-	13.4	-	-	-
SW1991/111	86	19	31.8	0.2	-	-	23	-	-	-
SW1991/112	90	32.9	54.8	0.71	0.088	0.36	11.2	3.4	6	11.7
SW1991/116	82	15.4	26.7	0.24	-	-	5	-	-	-
SW1991/117A	85	3.13	5.78	0.58	0.13	0.063	3.4	1.2	1.4	1.1
SW1991/120	73	38.2	63.5	1.2	-	-	9.2	-	-	-
SW1991/128	92	4.46	7.41	0.39	0.062	0.08	3	0.91	0.86	1.4
SW1991/135	73	10.4	18.1	0.46	-	-	4.5	-	-	-
SW1991/136	82	52.9	89.8	0.46	-	-	7.7	-	-	-
SW1991/142	83	4.45	7.56	0.1	-	-	0.58	-	-	-
SW1992/7	88	10.4	16.4	0.34	-	-	1.5	-	-	-

Table 8. Concentrations of chlorobiphenyl congeners in blubber (mg kg⁻¹ wet weight)

ReferenceNo.	%HEL	CB18	CB31	CB28	CB52	CB49	CB47	CB44	CB66	CB101	CB110	CB151	CB149	CB118
H. grypus														
SS1991/23	41	nd	nd	nd	0.23	0.08	0.21	nd	0.15	0.62	0.31	0.089	1.3	0.2
D. delphis														
SMRU90-73	91	nd	nd	0.12	0.35	0.064	nd	0.1	0.74	0.56	0.13	0.3	0.8	0.66
IC1990/4	85	0.043	0.026	nd	0.77	0.049	0.23	0.039	1.8	0.71	0.35	1.9	4.4	1.4
IC1991/1	75	nd	nd	nd	0.062	0.003	0.019	nd	nd	0.08	0.17	0.091	0.21	0.086
SW1990/5A	71	nd	nd	nd	0.55	nd	0.15	nd	nd	0.98	nd	1.2	2.8	1.1
SW1990/20A	55	0.023	0.027	nd	0.51	0.06	0.15	0.027	1.1	0.59	0.24	1.2	2.8	1
SW1990/45	75	nd	nd	nd	0.47	nd	0.1	nd	nd	1.1	0.12	0.4	1	0.65
SW1990/46	90	0.13	0.068	0.26	0.86	0.3	0.32	0.23	1.2	1.1	0.8	0.41	1.2	1.1
SW1990/47	82	nd	nd	0.15	0.44	0.16	0.19	0.14	0.28	0.5	5.1	0.14	0.45	0.44
SW1990/68	85	nd	nd	nd	2.2	nd	0.62	nd	nd	3.1	2	3.6	7.7	3.3
SW1990/78	86	0.065	0.029	nd	1.3	0.045	0.42	0.057	2.9	0.89	0.62	3.4	7.1	1.6
SW1990/81	82	nd	nd	nd	0.069	0.003	0.023	nd	nd	0.11	0.009	0.097	0.24	0.13
SW1990/87	88	nd	nd	nd	0.57	0.084	0.15	nd	nd	1.3	3.3	0.88	2.5	1.1
SW1990/88	78	0.021	nd	0.091	0.43	0.17	0.11	0.05	0.85	1.1	0.18	0.4	1.3	1.6
SW1990/97	79	0.087	0.096	nd	1.9	0.13	0.63	0.07	3.7	1.9	0.57	3.2	7.5	2.9
SW1990/101	82	0.031	0.036	nd	0.63	0.06	0.19	0.041	1.4	0.94	0.025	1.3	3.3	1.3
SW1990/102	82	0.026	0.028	nd	0.5	0.049	0.15	0.03	1.2	0.85	0.23	1.2	3	1.3
SW1990/103	80	0.029	0.047	nd	0.82	0.06	0.26	0.034	1.8	0.81	0.36	2	4.6	1.7
SW1990/104	80	0.03	0.028	nd	0.66	0.058	0.19	0.053	1.5	1.3	0.27	1.3	3.5	1.5
SW1990/108	91	0.03	0.06	nd	0.59	0.076	0.17	0.043	1.3	0.5	0.26	1.3	3.2	1.1
SW1991/7	87	nd	nd	nd	0.024	0.041	0.066	0.02	nd	0.15	0.12	0.14	0.4	0.25
SW1991/10	86	nd	nd	nd	2	nd	0.6	nd	nd	2.6	nd	2.8	5.9	2.6
SW1991/37	78	nd	nd	nd	1	0.16	0.29	nd	2.3	1.1	0.28	1.3	3.7	1.4
SW1991/83	90	nd	nd	nd	0.077	0.071	nd	nd	0.14	0.26	0.22	0.21	0.61	0.36
SW1991/137	76	0.024	0.015	nd	0.29	0.039	0.1	0.038	0.96	0.45	0.17	0.98	2.3	0.77
SW1992/1	82	0.056	nd	nd	1.8	0.32	0.56	0.049	2	1.4	0.49	3.3	7.5	1.9
SW1992/2	80	nd	nd	nd	0.91	0.21	0.27	nd	1.1	0.87	0.24	1.4	3.7	0.97
SW1992/3	84	nd	nd	nd	1.3	0.22	0.38	0.031	1.5	1.3	0.32	2	4.9	1.5
SW1992/4	84	nd	nd	nd	0.91	0.17	0.26	nd	1	0.75	0.26	1.4	3.6	1.1
SW1992/10	92	nd	nd	nd	0.75	0.25	0.22	nd	1	0.89	0.25	1.5	4	1.1
SW1992/11	88	nd	nd	nd	0.39	0.43	0.11	0.03	0.58	0.83	0.14	0.68	2	1.2
SW1992/12	64	nd	nd	nd	0.48	0.15	0.14	nd	0.62	0.39	0.2	1.2	2.4	0.59
SW1992/14(1)	90	nd	nd	nd	0.32	0.27	0.09	nd	0.5	0.38	0.13	0.79	1.9	0.71
SW1992/14(2)	92	nd	nd	nd	0.28	0.22	0.07	nd	0.41	0.49	0.09	0.51	1.4	0.58
SW1992/19(1)	84	nd	nd	nd	0.33	0.34	0.1	nd	0.49	0.61	0.11	0.58	1.6	0.94
SW1992/19(2)	86	nd	nd	nd	0.47	0.14	0.12	nd	0.6	0.51	0.14	0.83	2	0.76
G. griseus														
SW1990/70	88	nd	nd	nd	0.19	0.094	0.057	nd	0.28	0.3	0.12	0.22	0.76	0.48
G. melas														
SW1990/88A	84	nd	nd	nd	0.83	nd	0.23	nd	nd	1.4	nd	1.9	4.5	1.3
SW1991/55	75	nd	nd	0.066	1.3	0.34	0.21	0.08	1.4	2.1	0.14	0.93	3	2.4
L. acutus														
SW1989/88B	77	0.074	nd	0.14	1.2	0.085	0.33	0.083	3	0.098	0.24	1.2	3.1	0.78
S. coeruleoalba														
SW1991/26	56	nd	nd	0.061	0.47	0.027	0.14	0.028	1.2	0.62	0.16	0.67	1.7	0.67
T. truncatus														
SW1991/42	68	nd	nd	nd	2.5	0.089	0.9	nd	2.6	0.99	0.62	3.9	9.4	1.5
SW1991/115	57	nd	nd	nd	0.35	0.16	0.07	0.06	0.41	0.79	0.2	0.31	0.88	0.91
P. phocoena														
IC1990/1	84	nd	nd	nd	0.42	nd	0.099	nd	nd	0.61	nd	0.39	1.2	0.61
IC1990/2	73	nd	nd	nd	0.059	nd	0.011	nd	nd	0.027	nd	0.044	0.15	0.036
IC1990/3	70	nd	nd	nd	0.48	0.15	0.12	nd	nd	0.96	nd	0.56	1.6	0.97
IC1990/4	93	nd	nd	nd	0.26	nd	nd	nd	nd	0.23	nd	0.21	0.68	0.36
IC1991/2	62	0.017	nd	0.026	0.19	0.085	0.064	0.029	0.39	0.37	0.15	0.17	0.57	0.56
SMRU90-01	86	nd	nd	nd	0.53	0.064	0.054	nd	0.76	0.12	0.11	0.23	0.92	0.26
SMRU90-04	93	nd	nd	nd	0.16	0.11	nd	nd	0.22	0.28	0.14	0.11	0.34	0.28
SMRU90-02	89	nd	nd	nd	0.014	nd	nd	nd	nd	0.015	nd	0.01	0.028	0.014
SMRU90-03	82	nd	nd	nd	0.47	0.071	0.068	nd	0.64	0.089	0.087	0.23	0.88	0.18
SMRU90-05	87	nd	nd	nd	0.42	nd	nd	nd	nd	0.42	nd	0.22	0.65	0.25
SMRU90-06	89	nd	nd	nd	0.38	nd	nd	nd	nd	0.38	nd	0.21	0.69	0.27
SMRU90-07	83	nd	nd	nd	0.007	nd	nd	nd	nd	0.008	nd	0.006	0.017	0.006
SMRU90-08	91	nd	nd	nd	0.37	nd	nd	nd	nd	0.44	nd	0.22	0.64	0.35
SMRU90-09	87	nd	nd	nd	0.4	nd	nd	nd	nd	0.55	nd	0.28	0.85	0.44
SMRU90-10	72	nd	nd	nd	0.18	0.09	0.042	nd	0.3	0.076	nd	0.11	0.33	0.15
SMRU90-11	90	nd	nd	nd	0.008	nd	nd	nd	nd	0.009	nd	0.004	0.014	0.007
SMRU90-12	87	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd
SMRU90-13	76	nd	nd	nd	0.35	nd	nd	nd	nd	nd	nd	0.21	0.74	0.28
SMRU90-14	89	nd	nd	nd	0.51	0.076	0.09	nd	nd	0.67	nd	0.3	0.9	0.56
SMRU90-15	92	nd	nd	nd	0.65	nd	nd	nd	nd	nd	nd	0.32	1	0.2

Table 8. (continued)

ReferenceNo.	%HEL	CB18	CB31	CB28	CB52	CB49	CB47	CB44	CB66	CB101	CB110	CB151	CB149	CB118
P. phocoena														
SMRU90-16	91	nd	nd	nd	0.02	nd	nd	nd	nd	0.017	nd	0.01	0.03	0.01
SMRU90-17	87	nd	nd	nd	0.93	nd	nd	nd	nd	nd	nd	0.5	1.8	0.29
SMRU90-18	93	nd	nd	nd	1	nd	nd	nd	nd	nd	nd	0.58	2.1	0.33
SMRU90-19	89	nd	nd	nd	0.74	nd	nd	nd	nd	nd	nd	0.35	1.1	0.3
SMRU90-51	93	nd	nd	nd	0.27	0.12	0.081	0.049	0.42	0.5	0.2	0.49	1.4	0.65
SMRU90-55	85	0.067	nd	nd	3.6	0.26	0.52	0.074	5.7	1.7	0.85	2.4	7.8	2.7
SMRU90-60	92	nd	nd	nd	0.008	nd	nd	nd	nd	0.011	nd	0.004	0.015	0.007
SMRU90-61	89	nd	nd	nd	0.88	nd	0.11	nd	nd	nd	nd	0.58	2.3	0.34
SMRU90-75	93	nd	nd	nd	0.75	0.14	0.12	nd	1.3	0.35	0.11	0.44	1.4	0.43
SMRU90-76	89	nd	nd	nd	0.93	0.092	0.15	nd	nd	0.86	nd	0.66	2.2	0.49
SW1990/39	92	nd	nd	nd	0.52	0.09	0.14	nd	nd	0.5	nd	0.7	2.3	0.75
SW1990/50	95	nd	nd	nd	0.36	nd	0.11	nd	nd	0.27	nd	0.41	1.3	0.46
SW1990/50A	79	nd	nd	nd	0.74	nd	0.08	nd	nd	0.27	nd	0.29	1.2	0.34
SW1990/51(1)	78	nd	nd	nd	nd	0.092	nd	nd	0.088	0.089	nd	0.09	0.23	0.16
SW1990/51(2)	70	nd	nd	nd	nd	0.088	nd	nd	0.085	0.078	nd	0.063	0.18	0.14
SW1990/54	64	0.03	nd	nd	0.53	0.16	0.14	0.054	1.3	1	0.24	0.72	2.2	1.3
SW1990/65	76	nd	nd	nd	1.7	0.09	0.25	nd	nd	0.84	nd	1	3.6	0.85
SW1990/66	85	nd	nd	nd	0.75	0.089	0.18	nd	nd	0.83	1.2	0.77	2.6	1.5
SW1990/71	88	nd	nd	nd	0.06	0.066	nd	nd	0.1	0.073	nd	0.075	0.25	0.12
SW1990/72	81	0.06	nd	nd	0.6	0.096	0.15	nd	0.94	0.43	0.23	0.76	2.7	0.67
SW1990/72A	88	nd	nd	nd	0.19	0.094	0.057	nd	0.28	0.3	0.12	0.22	0.76	0.48
SW1990/73	71	0.046	nd	nd	1.2	0.084	0.2	0.035	2.1	0.51	0.3	0.84	3	0.91
SW1990/77B	85	0.046	nd	nd	1.5	0.065	0.21	0.042	3.1	0.39	0.48	1.8	4.7	nd
SW1990/79	87	nd	nd	nd	0.18	0.092	0.047	nd	0.25	0.26	0.085	0.13	0.44	0.33
SW1990/84	91	nd	nd	nd	1.3	0.12	0.12	nd	1.3	0.26	0.26	0.71	3.1	0.44
SW1990/92	83	nd	nd	nd	0.012	nd	nd	nd	nd	0.015	0.005	0.013	0.037	0.019
SW1990/94	75	0.03	nd	nd	1	0.07	0.12	0.04	1.7	0.37	0.24	0.77	2.6	0.5
SW1990/96	85	0.07	0.028	nd	2.1	0.888	0.26	0.059	3.2	0.51	0.64	2.1	8	1.5
SW1990/98	85	nd	nd	nd	0.089	0.007	0.014	nd	nd	0.056	0.006	0.066	0.19	0.064
SW1990/100	76	nd	nd	nd	0.057	0.003	0.009	nd	nd	0.03	nd	0.043	0.14	0.039
SW1990/100A	89	nd	nd	nd	0.9	0.1	0.14	nd	0.94	0.58	0.12	0.5	1.7	0.8
SW1990/106	76	0.061	nd	nd	0.89	0.19	0.23	0.042	1.6	1.3	0.39	1	3	2
SW1990/107	87	nd	nd	nd	0.15	nd	nd	nd	nd	0.25	nd	0.26	0.67	0.3
SW1990/109	89	0.27	0.021	nd	0.48	0.067	0.12	0.032	1.1	0.35	0.22	0.75	2.5	0.91
SW1990/111	93	nd	nd	nd	0.007	nd	nd	nd	nd	0.012	nd	0.013	0.033	0.015
SMRU91-06	92	nd	nd	nd	0.15	0.072	0.034	nd	0.28	0.18	0.036	0.11	0.39	0.18
SMRU91-08	90	nd	nd	nd	0.03	0.032	nd	nd	0.083	0.059	0.026	0.039	0.12	0.09
SMRU91-15	92	0.027	nd	0.015	0.73	0.08	0.13	0.019	1.2	0.43	0.16	0.53	1.9	0.77
SMRU91-07	93	nd	nd	nd	0.13	0.042	0.028	nd	0.23	0.13	0.034	0.13	0.45	0.18
SW1991/14	64	0.097	0.036	nd	2.8	0.14	0.29	0.08	3.8	0.81	0.82	2.8	9.6	1.5
SW1991/17	89	0.058	nd	nd	2	0.091	0.18	0.038	2.8	0.43	0.4	1.3	5.1	0.62
SW1991/17A	87	0.056	nd	nd	1.8	0.086	0.21	0.034	2.4	0.47	0.4	1.1	4.2	0.75
SW1991/19A	90	0.053	nd	0.027	1.6	0.092	0.16	0.32	2.3	0.37	0.34	1.1	4	0.059
SW1991/19B	89	0.021	nd	nd	0.28	0.052	0.065	0.021	0.67	0.29	0.13	0.45	1.4	0.56
SW1991/20	87	0.033	nd	nd	0.53	0.076	0.1	0.018	0.93	0.47	0.13	0.44	1.5	0.6
SW1991/22	85	0.016	nd	nd	0.77	0.049	0.081	nd	1.2	0.18	0.19	0.57	2.1	0.28
SW1991/23	86	0.058	nd	nd	2.6	0.13	0.34	0.027	3.7	1	0.51	1.8	6	1.2
SW1991/24	91	0.065	nd	0.027	1.7	0.17	0.22	0.037	2.4	0.52	0.4	1.2	4.3	0.77
SW1991/28	89	nd	nd	nd	0.39	0.071	0.058	nd	0.61	0.21	0.063	0.22	0.73	0.32
SW1991/29	88	0.016	nd	0.022	0.63	0.059	0.077	nd	0.86	0.24	0.083	0.3	1.1	0.37
SW1991/30	92	0.018	nd	0.038	0.22	0.074	0.051	nd	0.52	0.36	0.063	0.21	0.72	0.42
SW1991/31	87	0.027	nd	nd	1.1	0.11	0.19	0.022	2.2	0.7	0.35	1.3	4.2	1.3
SW1991/32A	91	nd	nd	nd	0.23	0.037	0.054	0.019	0.52	0.22	0.099	0.34	1	0.47
SW1991/35	74	nd	nd	nd	0.22	0.13	0.059	nd	0.36	0.31	0.064	0.19	0.49	0.58
SW1991/36	93	0.03	nd	nd	0.46	0.072	0.11	0.021	0.86	0.34	0.15	0.4	1.2	0.57
SW1991/43	88	nd	nd	nd	0.11	0.039	0.028	0.016	0.19	0.16	0.055	0.09	0.24	0.24
SW1991/48	86	nd	nd	nd	0.21	nd	nd	nd	0.34	nd	0.27	0.77	0.46	0.46
SW1991/54	90	nd	nd	nd	0.58	0.15	0.12	nd	0.68	0.97	0.14	0.6	1.8	1.1
SW1991/59	77	nd	nd	nd	0.18	0.2	0.059	nd	0.48	0.43	0.22	0.17	0.59	0.47
SW1991/61	52	nd	nd	nd	nd	0.13	nd	nd	0.29	0.2	nd	nd	0.39	nd
SW1991/63	75	nd	nd	nd	0.46	0.046	0.085	0.02	0.76	0.25	0.094	0.26	0.86	0.32
SW1991/70	92	nd	nd	nd	0.46	0.05	0.076	nd	0.79	0.27	0.24	0.68	1.9	0.79
SW1991/71	81	0.031	nd	0.032	0.55	0.1	0.14	0.026	1	0.63	0.17	0.54	1.7	1.1
SW1991/73	53	nd	nd	nd	0.27	0.19	0.078	0.04	0.62	0.54	0.2	0.24	0.83	0.51
SW1991/74	77	nd	nd	nd	0.084	0.096	0.04	nd	nd	0.21	0.14	0.14	0.46	0.3
SW1991/95	84	nd	nd	nd	0.54	0.075	0.12	0.022	0.93	0.49	0.16	0.54	1.7	0.81
SW1991/100	62	0.018	nd	nd	0.45	0.033	0.062	nd	0.67	0.16	0.078	0.22	0.86	0.2
SW1991/104	86	0.045	nd	nd	2.4	0.17	0.37	0.027	3.7	0.79	0.53	1.8	5.9	1.3
SW1991/111	86	nd	nd	nd	0.48	0.1	0.09	0.04	0.72	1.1	0.17	1	3.2	1.3
SW1991/112	90	0.027	nd	nd	2.1	0.064	0.26	0.018	2.5	0.37	0.44	1.2	5.1	0.69
SW1991/116	82	nd	nd	nd	0.81	0.15	0.15	nd	1.4	0.29	0.25	0.71	2.5	0.58
SW1991/117A	85	0.045	nd	0.028	0.34	0.088	0.064	0.036	0.65	0.37	0.061	0.2	0.52	0.42
SW1991/120	73	nd	nd	nd	2.3	0.28	0.32	nd	3.1	0.74	0.61	1.7	6.8	1.4
SW1991/128	92	nd	nd	nd	0.27	0.059	0.049	nd	0.52	0.22	0.063	0.2	0.66	0.34
SW1991/135	73	0.036	nd	nd	0.87	0.13	0.19	0.03	1.1	0.44	0.13	0.5	1.7	0.71
SW1991/136	82	nd	nd	nd	2.4	0.071	0.19	nd	3	0.37	0.79	1.6	8.1	nd
SW1991/142	83	nd	nd	nd	0.13	0.05	0.04	nd	0.26	0.16	0.066	0.19	0.62	0.27
SW1992/7	88	nd	nd	nd	0.51	0.064	0.086	nd	nd	0.18	0.14	0.43	1.4	0.43

Table 9. Concentrations of chlorobiphenyl congeners in blubber (mg kg⁻¹ wet weight)

Reference No.	% HEL	CB153	CB105	CB141	CB138	CB158	CB187	CB183	CB128	CB156	CB180	CB170	CB194
H. grypus													
SS1991/23	41	17.8	0.37	0.047	11.5	0.27	5	2.4	1.2	1	8.2	3.7	1
D. delphis													
SMRU90-73	91	1.5	0.22	nd	1.4	0.075	0.66	0.23	0.23	0.13	0.72	0.3	0.046
IC1990/4	85	10.8	1.4	0.067	8.8	0.47	3.6	1.3	1.3	0.88	4.4	2.2	0.32
IC1991/1	75	0.49	nd	0.01	0.43	0.031	0.21	0.068	0.055	nd	0.2	0.1	0.02
SW1990/5A	71	5.4	nd	0.16	4.6	0.27	2.3	0.78	0.65	0.21	2.4	1.2	0.14
SW1990/20A	55	7.7	0.86	0.05	5.6	0.27	2.8	0.97	0.62	0.59	3.4	1.3	1.27
SW1990/45	75	1.7	nd	0.06	1.6	0.1	0.84	0.23	0.25	0.06	0.57	0.24	0.043
SW1990/46	90	2.4	0.31	0.19	2	0.16	0.65	0.25	0.32	0.18	0.7	0.3	0.038
SW1990/47	82	0.91	0.14	0.09	0.73	0.076	0.29	0.11	0.11	0.055	0.29	0.12	0.031
SW1990/68	85	19.4	nd	nd	17.2	0.97	6.9	2.5	2.6	nd	3.8	7.3	0.6
SW1990/78	86	18.9	2.3	0.093	15.5	0.83	6.4	2.6	2.4	1.6	8.4	4	0.67
SW1990/81	82	0.48	nd	0.015	0.46	0.032	0.19	0.059	0.06	0.023	0.16	0.086	0.012
SW1990/87	88	5.3	nd	0.22	4.2	0.22	1.9	0.6	0.54	0.22	2	0.9	0.14
SW1990/88	78	3.2	0.41	0.083	2.8	0.17	0.73	0.29	0.41	0.23	1	0.46	0.055
SW1990/97	79	16.9	2.6	0.16	14.6	0.85	4.7	1.7	2.3	1.2	5.2	2.6	0.27
SW1990/101	82	7.7	1	0.11	6.1	0.32	2.6	0.89	0.8	0.61	3.1	1.4	0.22
SW1990/102	82	7.3	0.97	0.11	5.6	0.29	2.4	0.86	0.74	0.58	2.9	1.3	0.21
SW1990/103	80	11.8	1.4	0.08	9.6	0.5	3.8	1.4	1.1	0.89	4.5	2.1	0.26
SW1990/104	80	7.9	1	0.15	6.1	0.3	2.5	0.85	0.63	0.59	2.8	1.2	1.17
SW1990/108	91	8.1	1.1	0.043	6.7	0.33	2.7	1	0.91	0.66	3.7	1.6	0.26
SW1991/7	87	1.1	0.12	0.051	0.77	0.038	0.58	0.2	0.082	0.13	0.77	0.29	0.13
SW1991/10	86	12.7	nd	nd	12.2	0.85	4.7	1.5	2.4	nd	3.9	2.2	0.23
SW1991/37	78	7.4	1.4	nd	6	0.31	2.7	0.77	0.95	0.57	2.3	1.1	0.17
SW1991/83	90	1.8	0.089	0.077	1.2	0.063	0.88	0.29	0.18	0.12	1	0.41	0.2
SW1991/137	76	4.8	0.79	0.1	3.7	0.22	2	0.77	0.48	0.53	2.7	1.2	0.21
SW1992/1	82	17	1.2	0.11	14	0.72	5.7	1.9	2.2	0.72	5.5	2.6	0.38
SW1992/2	80	7.9	0.68	0.077	6.6	0.33	2.7	0.87	1	0.34	2.5	1.2	0.18
SW1992/3	84	11	0.88	0.11	9.1	0.49	3.9	1.3	1.5	0.49	3.6	1.7	0.27
SW1992/4	84	8.5	0.68	0.05	6.9	0.34	3	0.98	1	0.38	2.9	1.3	0.26
SW1992/10	92	9.2	0.69	0.093	7.2	0.34	3.3	1	1	0.4	3.3	1.4	0.23
SW1992/11	88	4.2	0.28	0.12	3.1	0.15	1.4	0.41	0.49	0.18	1.4	0.64	0.09
SW1992/12	64	8.1	0.44	0.025	5.8	0.27	3.1	1	0.77	0.35	3.3	1.4	0.31
SW1992/14(1)	90	4.7	0.35	0.029	3.5	0.17	1.6	0.52	0.51	0.21	1.7	0.78	0.12
SW1992/14(2)	92	3.3	0.27	0.06	2.4	0.12	1.2	0.39	0.36	0.15	1.3	0.52	0.09
SW1992/19(1)	84	3.3	0.21	0.07	2.6	0.13	1.1	0.38	0.4	0.16	1.1	0.54	0.076
SW1992/19(2)	86	4.8	0.4	0.035	3.8	0.19	1.5	0.53	0.57	0.2	1.6	0.72	0.097
G. griseus													
SW1990/70	88	2.5	0.19	0.06	2.1	0.11	0.7	0.25	0.25	0.13	0.92	0.36	0.06
G. melas													
SW1990/88A	84	11.4	nd	nd	9.4	0.48	4.4	1.5	0.13	nd	4.8	2.2	0.35
SW1991/55	75	5.4	0.51	0.11	4.3	0.27	1.6	0.64	0.63	0.26	2.2	0.86	0.16
L. acutus													
SW1989/88B	77	5.2	1.2	0.076	4.5	0.22	1.6	0.55	0.7	0.33	1.6	0.68	0.075
S. coeruleoalba													
SW1991/26	56	3.6	0.65	0.058	3.1	0.17	1.3	0.5	0.47	0.33	1.6	0.71	0.13
T. truncatus													
SW1991/42	68	31.5	1.5	0.067	21.5	1.4	10.8	4.4	3.2	1.5	14.1	5.2	1.6
SW1991/115	57	2.4	0.22	0.08	1.7	0.1	0.63	0.23	0.26	0.11	0.72	0.3	0.049
P. phocoena													
IC1990/1	84	1.5	1.3	0.087	1.3	0.087	0.53	0.16	0.17	0.21	0.44	0.21	0.032
IC1990/2	73	0.36	nd	nd	0.31	0.022	0.14	0.047	0.031	nd	0.13	0.063	0.012
IC1990/3	70	3.6	nd	0.17	3.1	0.17	1.4	0.4	0.41	nd	1.3	0.59	0.1
IC1990/4	93	1.9	nd	nd	1.6	0.1	0.82	0.26	0.22	nd	0.82	0.37	0.096
IC1991/2	62	1.6	0.22	0.06	1.3	0.078	0.39	0.14	0.18	0.12	0.41	0.18	0.035
SMRU90-01	86	2.8	0.38	nd	2.1	1.2	0.75	0.3	0.16	0.17	0.87	0.39	0.067
SMRU90-04	93	0.68	0.092	0.038	0.58	nd	0.17	0.05	0.08	0.07	0.18	0.064	nd
SMRU90-02	89	0.059	0.006	nd	0.048	0.003	0.022	0.006	0.005	nd	0.019	0.008	0.001
SMRU90-03	82	2.3	0.33	nd	1.8	0.11	0.69	0.26	0.12	0.16	0.73	0.35	0.055
SMRU90-05	87	1.4	nd	nd	1.3	0.07	0.41	0.14	0.13	nd	0.34	0.17	nd
SMRU90-06	89	1.3	nd	nd	1	nd	0.37	0.11	0.11	nd	0.28	0.14	nd
SMRU90-07	83	0.041	nd	nd	0.032	0.002	0.017	0.005	0.003	nd	0.015	0.006	0.001
SMRU90-08	91	1.4	nd	nd	1.2	0.069	0.46	0.14	0.13	nd	0.39	0.18	0.027
SMRU90-09	87	1.5	nd	nd	1.3	0.07	0.54	0.14	0.16	nd	0.39	0.18	0.024
SMRU90-10	72	0.9	0.14	nd	0.77	nd	0.21	0.085	0.087	nd	0.26	0.13	nd
SMRU90-11	90	0.031	nd	nd	0.024	nd	0.009	0.003	0.003	nd	0.007	0.003	nd
SMRU90-12	87	0.27	nd	nd	0.22	nd	nd	nd	nd	nd	0.16	nd	nd
SMRU90-13	76	1.4	nd	nd	1	nd	0.42	nd	nd	nd	0.35	0.15	nd
SMRU90-14	89	1.8	nd	nd	1.6	0.09	0.57	0.18	0.2	nd	0.48	0.23	0.033
SMRU90-15	92	2.3	nd	nd	1.7	0.095	0.64	0.23	0.1	nd	0.66	0.29	0.047
SMRU90-16	91	0.063	nd	nd	0.053	0.003	0.021	0.006	0.005	nd	0.017	0.008	0.001

Table 9. (continued)

ReferenceNo.	% HEL	CB153	CB105	CB141	CB138	CB158	CB187	CB183	CB128	CB156	CB180	CB170	CB194
P. phocoena													
SMRU90-17	87	3.5	nd	nd	2.8	0.14	1.1	0.34	0.18	nd	0.93	0.43	0.065
SMRU90-18	93	4.1	nd	nd	3.4	0.17	1.2	0.4	0.2	nd	1.1	0.5	0.072
SMRU90-19	89	2.5	nd	nd	1.9	0.11	0.67	0.24	0.13	nd	0.68	0.31	0.048
SMRU90-51	93	4.5	0.34	0.08	3.7	0.17	3	0.98	0.43	0.42	3.3	1.6	0.38
SMRU90-55	85	16.6	3.2	0.27	13.9	0.96	4.3	1.7	1.5	1.4	4.8	2.3	0.29
SMRU90-60	92	0.03	nd	nd	0.023	nd	0.009	nd	0.003	nd	0.006	0.003	nd
SMRU90-61	89	4.4	nd	nd	3.5	0.21	1.6	0.64	0.23	nd	1.5	0.68	0.11
SMRU90-75	93	2.7	0.58	nd	2	0.13	0.75	0.25	0.21	0.17	0.73	0.3	0.041
SMRU90-76	89	3.6	nd	nd	2.8	0.2	1.2	0.41	0.27	nd	1	0.46	0.072
SW1990/39	92	4	nd	nd	3.4	0.21	2	0.71	0.38	nd	1.8	0.89	0.18
SW1990/50	95	3.8	nd	nd	3.3	0.22	1.7	0.68	0.41	nd	1.8	0.88	0.21
SW1990/50A	79	2.7	nd	nd	2.2	0.14	0.68	0.23	0.18	nd	0.62	0.29	nd
SW1990/51(1)	78	0.87	0.052	0.093	0.75	0.038	0.87	0.3	0.1	0.18	1.2	0.39	0.29
SW1990/51(2)	70	0.54	0.043	0.036	0.45	nd	0.19	0.061	0.064	0.042	0.23	0.091	0.021
SW1990/54	64	4.5	0.86	0.16	3.7	0.22	1.5	0.51	0.53	0.36	1.5	0.66	0.11
SW1990/65	76	8.8	nd	nd	7.6	0.52	3.6	1.2	0.77	nd	3.4	1.8	0.36
SW1990/66	85	6	nd	nd	4.9	0.28	2	0.67	0.61	nd	2	0.94	0.16
SW1990/71	88	0.79	0.063	0.047	0.65	0.037	0.3	0.11	0.062	0.055	0.37	0.13	0.034
SW1990/72	81	7.7	0.82	0.063	6.4	0.34	3.6	1.2	0.68	0.51	3.2	1.5	0.22
SW1990/72A	88	2.5	0.19	0.06	2.1	0.11	0.7	0.25	0.25	0.13	0.92	0.36	0.06
SW1990/73	71	7.3	1.2	0.093	6	0.41	1.7	0.81	0.65	0.6	2.2	0.99	0.12
SW1990/77B	85	11.7	1.9	0.071	9.8	0.49	5.2	1.8	1.1	1.4	4.5	2.2	0.4
SW1990/79	87	0.96	0.12	0.032	0.84	0.043	0.25	0.087	0.11	0.048	0.27	0.1	0.016
SW1990/84	91	7.5	0.8	0.38	6.4	0.25	2.4	0.69	0.4	0.34	2	0.97	0.12
SW1990/92	83	0.058	nd	nd	0.051	0.003	0.04	0.012	0.006	nd	0.035	0.016	0.009
SW1990/94	75	5.4	0.96	0.03	4.3	0.24	2	0.63	0.33	0.43	1.7	0.8	0.17
SW1990/96	85	18.6	2.6	0.08	14.2	0.79	5.7	2.3	1.2	1.5	6.8	3.3	0.48
SW1990/98	85	0.42	nd	nd	0.36	0.023	0.14	0.044	0.033	nd	0.12	0.066	0.007
SW1990/100	76	0.31	nd	nd	0.27	0.017	0.11	0.034	0.026	nd	0.096	0.05	0.007
SW1990/100A	89	3.7	0.45	0.042	3.3	0.16	0.91	0.27	0.33	0.13	0.84	0.36	0.036
SW1990/106	76	8.2	1	0.18	6.5	0.38	2	0.79	0.92	0.58	2.3	1.1	0.17
SW1990/107	87	1.6	nd	nd	1.5	0.08	1.1	0.34	0.16	nd	0.99	0.47	0.18
SW1990/109	89	6.1	0.85	0.056	4.5	0.22	1.8	0.59	0.48	0.43	1.9	0.87	0.11
SW1990/111	93	0.084	nd	nd	0.076	0.004	0.056	0.017	0.007	nd	0.05	0.024	0.009
SMRU91-06	92	0.89	0.14	nd	0.76	0.025	0.27	0.082	0.079	0.067	0.22	0.096	0.025
SMRU91-08	90	0.42	0.053	nd	0.31	nd	0.18	0.069	0.035	0.05	0.23	0.091	0.028
SMRU91-15	92	5.2	0.68	0.046	5	0.22	1.4	0.47	0.38	0.34	1.4	0.59	0.097
SMRU91-07	93	1.1	0.12	nd	0.77	0.039	0.34	0.11	0.049	0.074	0.33	0.13	0.21
SW1991/14	64	22.1	nd	0.1	17.4	0.88	7.2	2.8	1.1	1.8	8.1	3.9	0.65
SW1991/17	89	10.4	nd	0.036	8.2	0.36	3	1	0.53	0.69	2.5	1.2	0.17
SW1991/17A	87	7.6	1.5	0.034	6	0.29	1.9	0.56	0.51	0.47	1.5	0.84	0.081
SW1991/19A	90	8	nd	0.027	6.2	0.3	2.2	0.71	0.44	0.52	1.9	0.87	0.12
SW1991/19B	89	3.3	0.52	0.05	2.6	0.15	1.1	0.37	0.29	0.26	1.1	0.5	0.075
SW1991/20	87	2.6	0.57	0.046	2.3	0.11	0.78	0.22	0.3	0.18	0.61	0.29	0.033
SW1991/22	85	4.9	nd	nd	3.8	0.16	1.5	0.51	0.2	0.34	1.4	0.6	0.11
SW1991/23	86	13.1	1.9	0.055	10	0.41	2.7	0.72	0.79	0.59	2.1	0.95	0.087
SW1991/24	91	8.8	1.6	0.038	7.4	0.33	2.5	0.79	0.55	0.59	2.2	1.1	0.14
SW1991/28	89	1.7	0.29	0.017	1.3	0.069	0.42	0.14	0.14	0.092	0.038	0.16	0.019
SW1991/29	88	2.4	0.43	0.019	2.1	0.099	0.51	0.16	0.18	0.14	0.46	0.22	0.023
SW1991/30	92	1.6	0.29	0.036	1.2	0.057	0.41	0.12	0.15	0.1	0.35	0.16	0.024
SW1991/31	87	9.8	1.4	0.049	7.8	0.35	2.4	0.67	0.69	0.52	2.1	1	0.11
SW1991/32A	91	2.6	0.39	0.04	1.9	0.1	0.86	0.28	0.22	0.2	0.95	0.42	0.065
SW1991/35	74	1.9	0.21	0.025	1.6	0.065	0.35	0.099	0.24	0.098	0.3	0.15	0.015
SW1991/36	93	2.5	0.51	0.031	2.2	0.11	0.73	0.21	0.29	0.19	0.63	0.35	0.072
SW1991/43	88	0.7	0.1	0.02	0.6	0.034	0.21	0.82	0.098	0.053	0.24	0.12	0.024
SW1991/48	86	2	nd	nd	1.7	0.076	0.9	0.27	0.21	nd	0.77	0.39	0.13
SW1991/54	90	4.1	0.39	0.097	3.2	0.15	1.2	0.34	0.41	0.16	1	0.47	0.08
SW1991/59	77	1.1	0.26	0.04	0.86	0.045	0.23	0.063	0.14	0.056	0.21	0.1	nd
SW1991/61	52	0.75	0.16	nd	0.56	nd	0.3	nd	nd	nd	0.27	nd	nd
SW1991/63	75	1.9	0.35	0.029	1.4	0.11	0.44	0.18	0.15	0.12	0.48	0.22	0.028
SW1991/70	92	7.5	0.72	0.018	6.4	0.2	2	0.41	0.42	0.45	1.2	0.69	0.075
SW1991/71	81	4	0.69	0.1	3.5	0.22	1.3	0.46	0.49	0.34	1.5	0.66	0.11
SW1991/73	53	1.9	0.33	0.067	1.4	0.094	0.48	0.17	0.19	0.13	0.53	0.22	0.029
SW1991/74	77	1	0.19	0.052	0.82	0.059	0.38	0.15	0.14	0.11	0.46	0.18	0.034
SW1991/95	84	4	0.53	0.04	3.1	0.17	1.1	0.36	0.37	0.24	1.2	0.49	0.08
SW1991/100	62	1.8	0.36	0.016	1.4	0.097	0.36	0.15	0.14	0.11	0.38	0.17	0.02
SW1991/104	86	12.8	2.2	0.065	10.8	0.7	4.3	1.6	0.93	1	4.4	1.9	0.36
SW1991/111	86	7	0.53	0.01	5.2	0.25	2.9	1	0.71	0.35	3.9	1.5	0.28
SW1991/112	90	14.5	1.4	0.036	10.5	0.62	4.6	1.7	0.43	1.1	4.7	2.1	0.35
SW1991/116	82	6.4	0.93	0.033	5.1	0.3	2.1	0.72	0.45	0.51	2.2	0.94	0.16
SW1991/117A	85	0.9	0.23	0.044	0.8	0.046	0.26	0.092	0.12	0.057	0.27	0.12	0.014
SW1991/120	73	16.7	2	0.09	12.8	0.64	4.2	1.4	1.1	0.86	4.3	1.9	0.25
SW1991/128	92	1.8	0.28	0.015	1.4	0.067	0.42	0.14	0.15	0.11	0.43	0.18	0.032
SW1991/135	73	4	0.64	0.37	3.4	0.19	1.2	0.4	0.36	0.23	1	0.44	0.079
SW1991/136	82	23.5	2.7	0.051	17.2	0.89	8.2	3.7	0.93	1.8	9.4	4	0.89
SW1991/142	83	1.8	0.21	0.025	1.4	0.063	0.68	0.24	0.15	0.15	0.69	0.31	0.055
SW1992/7	88	4.4	0.49	nd	3.4	0.17	1.3	0.51	0.3	0.32	1.5	0.65	0.13

Table 10. Concentrations of organochlorine compounds in stomach contents of common dolphins (mg kg⁻¹ wet weight)

ReferenceNo.	%HEL	ICES7 CBs	Total25CBs	HCB	α-HCH	γ-HCH	p,p'-DDE	p,p'-DDT	p,p'-TDE	Dieldrin
D.delphis										
SW1992/2	8	0.024	0.059	nd	0.005	0.001	0.009	0.007	0.017	0.007
SW1992/4	5	0.025	0.046	0.002	0.004	0.001	0.006	0.006	0.01	0.006
SW1992/19(1)	18	0.111	0.24	0.01	0.027	0.004	0.021	nd	0.037	0.042
SW1992/19(2)	3	0.017	0.038	nd	nd	nd	0.004	0.005	0.003	0.005
SW1992/20	2	0.013	0.029	nd	nd	nd	0.005	0.003	0.01	0.01

ReferenceNo.	% HEL	CB18	CB31	CB28	CB52	CB49	CB47	CB44	CB66	CB101	CB110	CB151	CB149	CB118
D.delphis														
SW1992/2	8	nd	nd	nd	nd	0.023	nd	nd	nd	0.002	nd	nd	0.004	0.002
SW1992/4	5	nd	nd	nd	nd	0.007	nd	nd	nd	0.002	0.003	0.001	0.004	0.003
SW1992/19(1)	18	nd	nd	nd	nd	0.053	nd	nd	nd	nd	0.012	0.007	0.018	0.013
SW1992/19(2)	3	nd	nd	nd	nd	0.012	nd	nd	nd	nd	nd	nd	0.003	0.002
SW1992/20	2	nd	nd	nd	nd	0.011	nd	nd	nd	nd	nd	nd	0.002	0.002

ReferenceNo.	% HEL	CB153	CB105	CB141	CB138	CB158	CB187	CB183	CB128	CB156	CB180	CB170	CB194
D.delphis													
SW1992/2	8	0.01	0.001	0.001	0.007	nd	0.004	nd	nd	nd	0.003	0.002	nd
SW1992/4	5	0.01	nd	nd	0.007	nd	0.003	nd	0.001	nd	0.003	0.002	nd
SW1992/19(1)	18	0.05	0.005	nd	0.031	nd	0.016	0.005	0.005	nd	0.017	0.008	nd
SW1992/19(2)	3	0.007	0.001	nd	0.005	nd	0.003	nd	0.001	nd	0.003	0.001	nd
SW1992/20	2	0.005	nd	nd	0.004	nd	0.002	nd	nd	nd	0.002	0.001	nd

This programme has also created the largest dataset to date on contaminants in marine mammal tissues collected using a single set of analytical methodology. Comparisons between individuals can therefore be made without the usual caveats concerning comparability of concentrations determined in different laboratories and by different methods (Wagemann and Muir, 1984; Law *et al.*, 1989(a) and (b)). Because both detailed pathological data and tissue contaminant concentrations were available from the same individuals, it was possible to investigate possible relationships between tissue levels of environmental contaminants and a number of pathological conditions. No evidence was found that chlorinated hydrocarbons result in an increased susceptibility to infectious and parasitic diseases in harbour porpoises. Porpoises that died from an infectious or parasitic disease were in significantly poorer body condition than porpoises that died from acute physical trauma (Kuiken *et al.*, 1994(b)). This may indicate that thinner porpoises are more susceptible to infections and parasitic diseases, or alternatively may reflect the fact that, once ill, animals lose appetite and so their body condition deteriorates. There was no correlation between blubber levels of chlorinated hydrocarbons and adrenocortical hyperplasia in harbour porpoises, but this lesion did occur in porpoises that died after prolonged illness (Kuiken *et al.*, 1993).

These findings increase our knowledge of causes of disease and death in marine mammals, and contribute to a better understanding of the lesions found on postmortem examination and how they relate to environmental

contamination. Among other things, this improves the diagnosis of causes of mortality in marine mammals. Marine mammals are subjected to a variety of potentially harmful factors in their environment, all of which can potentially interact with each other. To understand the effects they have on marine mammal populations, a multidisciplinary approach is essential. With this project we hope to have made a step in the right direction.

6. REFERENCES

- ALLCHIN, C. R., KELLY, C. A. AND PORTMANN, J. E. 1989. Methods of analysis for chlorinated hydrocarbons in marine and other samples. *Aquat. Environ. Prot.: Analyt. Meth.*, MAFF Direct. Fish. Res., Lowestoft, (6), 25pp.
- ANON. 1986. Report of the ICES Advisory Committee on Marine Pollution 1985. *ICES Coop. Res. Rep.*, 135, 82pp.
- BALLSCHMITER, K. AND ZELL, M. 1980. Analysis of polychlorinated biphenyls (PCB) by glass capillary gas chromatography. *Fresenius Z. Anal. Chem.*, 302, 20-31.
- BROUWER, A, REIJNDERS, P. J. H. AND KOEMAN, J. H. 1989. Polychlorinated biphenyl (PCB)-contaminated fish induces vitamin A and thyroid hormone deficiency in the common seal *Phoca vitulina*. *Aquat. Toxicol.*, 15, 99-106.

- DIETZ, R., HEIDE-JORGENSEN, M.-P. AND HÄRKÖNEN, T. 1989. Mass deaths of harbor seals (*Phoca vitulina*) in Europe. *Ambio*, **18**, 258-264.
- HARPER, D. J., FILEMAN, C. F., MAY, P. V. AND PORTMANN, J. E. 1989. Methods of analysis for trace metals in marine and other samples. *Aquat. Environ. Prot.: Analyt. Meth., MAFF Direct. Fish. Res., Lowestoft*, (3), 38pp.
- JONES, B. R. AND LASLETT, R. E. (IN PREP.). Methods of analysis for trace metals in marine and other samples. *Aquat. Environ. Prot.: Analyt. Meth., MAFF Direct. Fish. Res., Lowestoft*.
- KENNEDY, S., KUIKEN, T., ROSS, H. M., McALISKEY, M., MOFFETT, D., McNIVEN, C. M. AND CAROLE, M. 1992. Morbillivirus infection in two common porpoises (*Phocoena phocoena*) from the coasts of England and Scotland. *Veterinary Record*, **131**, 286-290.
- KUIKEN, T. AND BAKER, J. R. 1991(a). Guidelines for the postmortem and tissue sampling of cetaceans. London, Zoological Society of London, 16pp.
- KUIKEN, T. AND BAKER, J. R. 1991(b). Guidelines for the postmortem and tissue sampling of seals. London, Zoological Society of London, 16pp.
- KUIKEN, T. AND BAKER, J. R. 1993. Guidelines for the postmortem and tissue sampling of cetaceans. London, Zoological Society of London, 16pp.
- KUIKEN, T. AND GARCIA HARTMANN, M. (EDS.) 1993. Proceedings of the first European Cetacean Society workshop on cetacean pathology: dissection techniques and tissue sampling. Leiden, The Netherlands, 13-14 September 1991. *ECS Newsletter* 17, 39pp.
- KUIKEN, T., HOFLE, U., BENNETT, P. M., ALLCHIN, C. R., KIRKWOOD, J. K., BAKER, J. R., APPLEBY, E. C., LOCKYER, C. H., WALTON, M. J. AND SHELDRIK, M. C. 1993. Adrenocortical hyperplasia, disease and chlorinated hydrocarbons in the Harbour porpoise (*Phocoena phocoena*). *Mar. Pollut. Bull.*, **26**, 440-446.
- KUIKEN, T., SIMPSON, V. R., ALLCHIN, C. R., BENNETT, P. M., CODD, G. A., HARRIS, E., HOWES, G. J., KENNEDY, S., KIRKWOOD, J. K., LAW, R. J., MERRETT, N. R. AND PHILLIPS, S. 1994(a). Mass mortality of common dolphins (*Delphinus delphis*) in south-west England due to incidental capture in fishing gear. *Veterinary Record*, **134**, 81-89.
- KUIKEN, T., BENNETT, P. M., ALLCHIN, C. R., KIRKWOOD, J. K., BAKER, J. R., LOCKYER, C. H., WALTON, M. J. AND SHELDRIK, M. C. 1994(b). PCB's, cause of death and body condition in harbour porpoises (*Phocoena phocoena*) from British Waters. *Aquat. Toxicol.*, **28**, 13-28.
- LAW, R. J., ALLCHIN, C. R. AND HARWOOD, J. 1989(a). Concentrations of organochlorine compounds in the blubber of seals from eastern and north-eastern England, 1988. *Mar. Pollut. Bull.*, **20**, 110-115.
- LAW, R. J., ALLCHIN, C. R. AND HARWOOD, J. 1989(b). Reply to letter by Simmonds and Johnston. *Mar. Pollut. Bull.*, **20**, 583-584.
- LOCKYER, C. (IN PRESS). A review of factors involved in zonation in odontocete teeth, and an investigation of the likely impact of environmental factors and major life events on harbour porpoise tooth structure. *Rep. int. Whal. Comm. (Special Issue)*.
- MORRIS, R. J., LAW, R. J., ALLCHIN, C. R., KELLY, C. A. AND FILEMAN, C. F. 1989. Metals and organochlorines in dolphins and porpoises of Cardigan Bay, West Wales. *Mar. Pollut. Bull.*, **20**, 512-523.
- OSTERHAUS, A. D. M. E., BROEDERS, H. W. J., TEPPEMA, J. S., KUIKEN, T., HOUSE, J. A., VOS, H. W. AND VISSER, I. K. G. 1993. Isolation of a virus with rhabdovirus morphology from a white-beaked dolphin (*Lagenorhynchus albirostris*). *Archives of Virology*, **133**, 189-193.
- SIMMONDS, M. P. AND JOHNSTON, P. A. 1989. Seals, sense and science (1). *Mar. Pollut. Bull.*, **20**, 580-583.
- VISSER, L. K. G., VAN BRESSEM, M.-F., DE ZWART, H. L., VAN DE BILDT, M. W. G., VOS, H. W., VAN DER HEIJDEN, R. W. J., SALIKI, J. T., ORVELL, C., KITCHING, P., KUIKEN, T., BARRETT, T. AND OSTERHAUS, A. D. M. E. 1993. Characterisation of morbilliviruses isolated from dolphins and harbour porpoises in Europe. *Journal of General Virology*, **74**, 631-641.
- WAGEMANN, R. AND ARMSTRONG, F. A. J. 1988. Trace metal determination in animal tissues: an interlaboratory comparison. *Talanta*, **35**, 545-551.
- WAGEMANN, R. AND MUIR, D. C. G. 1984. Concentrations of heavy metals and organochlorines in marine mammals of northern waters: overview and evaluation. *Can. Tech. Rep. Fish. Aquat. Sci.*, **1279**, 97pp.

7. ACKNOWLEDGEMENTS

This project was partially funded by the Department of the Environment as part of its co-ordinated programme of research on the North Sea. MAFF and SOAFD contributed analytical resources under their own programmes of environmental research. We would like to thank Charlotte Iskjaer-Ackley for her considerable efforts in preparation of the main data tables in this report whilst working at the Institute of Zoology under a

contract (POSEIDON: UK central database for marine mammal strandings PECD 7/8/247) from the Department of the Environment. We would also like to thank the MAFF Veterinary Investigation Service and the Scottish Agricultural College for assisting with this work and are particularly grateful to all the veterinary surgeons who undertook postmortem investigations: Mr W. L. G. Ashton, Dr J. R. Baker, Mr J. Barker, Mr C. H. G. Bishop, Mr A. C. Bygrave, Mr A. J. Cook, Mr M. P. Cranwell, Mr A. A. Cunningham, Mr I. H. Davies, Dr F. M. D. Gulland, Mr M. Garcia Hartmann, Mr D. G. Harwood, Mr F. D. Kirby, Dr J. K. Kirkwood, Mr T. Kuiken, Ms M. T. O'Leary, Mr I. Robinson, Mr V. R. Simpson, Mr D. P. Stubbings and Mr B. W. Tatton: and would also like to thank Dr C. H. Lockyer and Dr A. W. Martin for organising annual harbour porpoise postmortem sessions at SMRU. We are also grateful to the many individuals and organisations who assisted the work by reporting strandings, transporting carcasses, and by contributing in many other ways.

8. PARTICIPATING ORGANISATIONS

IOZ :
Institute of Zoology
Regents Park
London NW1 4RY

MAFF :
MAFF Fisheries Laboratory
Remembrance Avenue
Burnham-on-Crouch
Essex CM0 8HA

NHM :
Natural History Museum
Cromwell Road
London SW7 5BD

SMRU :
Sea Mammal Research Unit
Natural Environment Research Council
High Cross, Madingley Road
Cambridge CB3 0ET

SOAFD :
Scottish Office Agriculture and Fisheries Department
Marine Laboratory
P O Box 101
Victoria Road
Aberdeen AB9 8DB

UL :
University of Liverpool
Veterinary Field Station
Neston, Wirral L64 7TE

Present Address :
Thijs Kuiken
Department of Veterinary Pathology
Western College of Veterinary Medicine
University of Saskatchewan
Saskatoon, Saskatchewan S7N 0W0
Canada

The reference to proprietary products in this report should not be construed as an official endorsement of these products, nor is any criticism implied of similar products which have not been mentioned.

APPENDIX 1. Cetacean postmortem report form

CETACEAN POSTMORTEM REPORT

SW NO.:..... PM NO.:..... OTHER ID NO.:

SPECIES:..... SEX:

LOCATION FOUND:..... DATE FOUND:.....

FOUND BY (name, address):

.....

PATHOLOGIST:..... DATE OF PM:.....

FROZEN?: Y/N

1. BASIC MEASUREMENTS

BODY CONDITION USING CONDITION CODE:.....

Condition code:

- 1) **live** (becomes code 2 at death)
- 2a) **extremely fresh** (as if just died, no bloating, meat is considered by most to be edible)
- 2b) **slight decomposition** (slight bloating, blood imbibition visible)
- 3) **moderate decomposition** (moderate bloating, skin peeling, penis may be extended in males, organs still intact, excluding postmortem damage)
- 4) **advanced decomposition** (major bloating, skin peeling, penis extended in males, organs beyond recognition, bones exposed due to decomposition)
- 5) **indeterminate** (mummified carcass or skeletal remains, no organs present)

PHOTOGRAPHS TAKEN

-lateral views whole body, both sides: Y/N

-lateral views dorsal fin, both sides: Y/N

-baleen whales: ventral view tail flukes: Y/N

-other photographs (list):

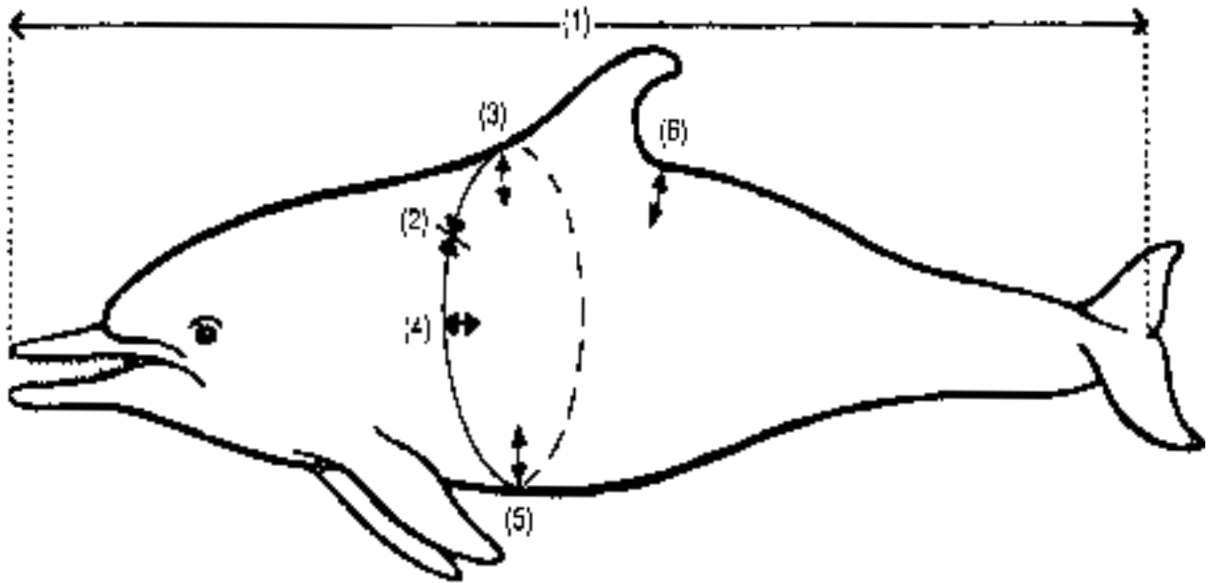
- -

- -

BODY WEIGHT (kg):.....

LENGTH, GIRTH AND BLUBBER THICKNESS (see diagram below):

- tip upper jaw to tail notch (cm) (1):
- girth in front of dorsal fin (cm) (2):
- blubber thickness in front of dorsal fin:
- dorsal mid-line (mm) (3):
- lateral (mm) (4):
- ventral mid-line (mm) (5):
- blubber thickness behind dorsal fin:
- dorsal mid-line (mm) (6):



2. GROSS PATHOLOGICAL EXAMINATION

*Encircle the appropriate category: NE = not examined
NAD = nothing abnormal detected
A = abnormal (describe fully in section 5)*

EXTERNAL EXAMINATION

NE NAD A -body orifices
NE NAD A -fins and flukes
nutritional state: good / moderate / poor

INTEGUMENT

NE NAD A -epidermis
NE NAD A -blubber
NE NAD A -subcutaneous tissue
NE NAD A -mammary glands

MUSCULOSKELETAL SYSTEM

NE NAD A -skull
NE NAD A -other bones
NE NAD A -back muscle mass
NE NAD A -other muscles

NERVOUS SYSTEM

NE NAD A -brain
NE NAD A -spinal cord
NE NAD A -peripheral nerves

CARDIOVASCULAR SYSTEM

NE NAD A -pericardial sac
NE NAD A -myocardium
NE NAD A -valves
NE NAD A -arteries, veins

RESPIRATORY SYSTEM

NE NAD A -nasal cavity
NE NAD A -sinuses
NE NAD A -trachea, bronchi
NE NAD A -lungs
NE NAD A -pleura/pleural cavity

ALIMENTARY SYSTEM

NE NAD A -mouth
NE NAD A -oesophagus
NE NAD A -cardiac section stomach
NE NAD A -fundic section stomach
NE NAD A -pyloric section stomach
NE NAD A -intestine
NE NAD A -anus
NE NAD A -liver
NE NAD A -pancreas
NE NAD A -peritoneum/
peritoneal cavity

UROGENITAL SYSTEM

NE NAD A -kidneys
NE NAD A -ureters
NE NAD A -urinary bladder
NE NAD A -urethra
NE NAD A -ovaries/testes
NE NAD A -uterus
NE NAD A -vagina/penis
NE NAD A -vulva/preputium

LYMPHATIC AND ENDOCRINE SYSTEMS

NE NAD A -adrenal glands
NE NAD A -thyroid gland
NE NAD A -spleen
NE NAD A -thymus
NE NAD A -lymph nodes

10% Formalin

<input type="checkbox"/>	adrenal glands	both
<input type="checkbox"/>	brain	whole
<input type="checkbox"/>	heart	1 cm ³
<input type="checkbox"/>	kidney	1 cm ³
<input type="checkbox"/>	liver	1 cm ³
<input type="checkbox"/>	lung	4 x 1 cm ³
<input type="checkbox"/>	lung (for Kennedy)	4 x 1 cm ³
<input type="checkbox"/>	mammary gland	1 cm slice
<input type="checkbox"/>	mesenteric ln.	1 cm slice
<input type="checkbox"/>	ovaries*	both
<input type="checkbox"/>	pancreas	1 cm ³
<input type="checkbox"/>	pulm. ass. ln.	1 cm slice
<input type="checkbox"/>	spleen	1 cm ³
<input type="checkbox"/>	testes*	both/slices
<input type="checkbox"/>	thymus	1 cm ³

Bacteriology

<input type="checkbox"/>	heart blood	-
<input type="checkbox"/>	kidney	swab/block
<input type="checkbox"/>	liver	swab/block
<input type="checkbox"/>	lung	swab/block

Virology (freeze at -70°C)

<input type="checkbox"/>	brain	1 cm ³
<input type="checkbox"/>	kidney	1 cm ³
<input type="checkbox"/>	lung	1 cm ³

4. LIST OF EXTRA SAMPLES

Extra samples of
lesions taken for
histological examination
(list):

-
-
-
-
-
-

Extra samples of
lesions taken for
bacteriological examination
(list):

-
-
-
-
-
-

Other extra samples
taken (list):

-
-
-
-
-
-
-

5. DESCRIPTION OF ABNORMALITIES GROSS PATHOLOGICAL EXAMINATION

(add extra pages if necessary)

PRELIMINARY DIAGNOSIS OF GROSS PATHOLOGICAL EXAMINATION (in order of importance):

a.

b.

c.

d.

e.

6. RESULTS OF HISTOLOGICAL EXAMINATION (add extra pages if necessary)

7. RESULTS OF BACTERIOLOGICAL EXAMINATION

Heart blood:

Lung:

Liver:

Kidney:

Other:

.

.

8. MISCELLANEOUS RESULTS

9. FINAL DIAGNOSIS (in order of importance):

a.

b.

c.

d.

e.

APPENDIX 2. Summary of results derived from the analysis of reference materials certified for trace metals (MAFF)

(a) Analyses conducted in 1991.

Units are mg kg⁻¹. Mean value ± one standard deviation given.

NRC* DOLT-1 Dogfish liver

	MAFF value	(n)	Certified value
Cd	3.90 ± 0.27	13	4.18 ± 0.28
Cu	18.5 ± 1.2	13	20.8 ± 1.2
Hg	0.25 ± 0.06	14	0.22 ± 0.03
Zn	86.4 ± 5.3	13	92.5 ± 2.3

n = number of analyses

NIST[§] 1566a Oyster tissue

	MAFF value	(n)	Certified value
Hg	0.06 ± 0.01	17	0.06 ± 0.01
Cd	3.8 ± 0.6	14	4.15 ± 0.38
Cu	60.0 ± 4.7	19	66.3 ± 4.3
Zn	810 ± 72	18	830 ± 57
Ni	1.85 ± 0.55	7	2.25 ± 0.44
Cr	1.95 ± 0.36	4	1.43 ± 0.46

NRC TORT-1 Lobster hepatopancreas

	MAFF value	(n)	Certified Value
Pb	10.3 ± 1.0	19	10.4 ± 2.0
Zn	168 ± 9.8	20	177 ± 10.0
Hg	0.34 ± 0.06	28	0.33 ± 0.06
Cd	24.1 ± 1.1	19	26.3 ± 2.1
Cu	408 ± 14	19	439 ± 22

NRC DORM-1 Dogfish muscle

	MAFF value	(n)	Certified value
Cu	4.97 ± 0.41	14	5.22 ± 0.30
Hg	0.63 ± 0.05	19	0.79 ± 0.07
Zn	19.2 ± 1.7	13	21.3 ± 1.0

IAEA[¶] MAA2/TM Fish muscle

	MAFF value	(n)	Certified value
Cu	3.85 ± 0.35	27	4.0 ± 0.10
Hg	0.41 ± 0.06	42	0.47 ± 0.02
Zn	30.8 ± 1.6	27	33.0 ± 1.0

Sources of CRMs :

* NRC - National Research Council, Canada

§ NIST - National Institute of Standards and Technology, USA

¶ IAEA - International Atomic Energy Agency, Monaco.

(b) Analyses conducted in 1992.

Units are mg kg⁻¹. Mean value ± one standard deviation given.

NBS 1566 a Oyster tissue

	MAFF value	(n)	Certified value
Cd	4.12 ± 0.28	37	4.15 ± 0.38
Cu	61.8 ± 3.3	36	66.3 ± 4.3
Hg	0.06 ± 0.01	39	0.06 ± 0.007
Zn	829 ± 53	37	830 ± 57
Se	2.29 ± 0.3	9	2.21 ± 0.24

NRC DOLT-1 Dogfish liver

	MAFF value	(n)	Certified value
Hg	0.24 ± 0.04	70	0.225 ± 0.037
Cd	4.31 ± 0.41	57	4.18 ± 0.28
Cu	19.3 ± 1.1	62	20.8 ± 1.2
Zn	85.2 ± 6.2	61	92.5 ± 2.3
Pb	1.18 ± 0.23	23	1.36 ± 0.29
Se	6.5 ± 0.4	12	7.34 ± 0.42

NRC TORT-1 Lobster hepatopancreas

	MAFF value	(n)	Certified Value
Pb	9.6 ± 13	21	10.4 ± 2.0
Zn	156 ± 10	16	177 ± 10.0
Hg	0.32 ± 0.06	17	0.33 ± 0.06
Cd	23.4 ± 1.6	19	26.3 ± 2.1
Se	6.36 ± 0.46	5	6.88 ± 0.47
Cu	390 ± 34	16	439 ± 22

NRC DORM-1 Dogfish muscle

	MAFF value	(n)	Certified value
Cr	3.12 ± 0.44	44	3.6 ± 0.40
Cu	5.12 ± 0.47	85	5.22 ± 0.30
Hg	0.69 ± 0.09	92	0.79 ± 0.07
Zn	19.2 ± 2.0	83	21.3 ± 1.00
Pb	0.35 ± 0.10	27	0.40 ± 0.12
Se	1.69 ± 0.28	17	1.62 ± 0.12