Annex A – Norovirus Sample Submission Instructions

Types and condition of shellfish accepted for testing:

Samples must be submitted as <u>either live or frozen shellfish</u>, and in all cases must be uncooked, unprocessed and unshucked.

The customer will be responsible for collecting the samples. Before submitting live shellfish, mud and sediment adhering to the shellfish should be removed (where possible). This is best achieved by rinsing/scrubbing with clean sea water or freshwater of potable quality. If these are not available, the seawater from the immediate area of sampling may be used instead. Do not totally re-immerse the shellfish in water as this may cause them to open. Allow them to drain before placing them in the sample container.

Samples should arrive at the laboratory in good condition (opened, gaping or damaged shells should not be included in the sample). Samples should only consist of animals that are within the normal commercial size range. Immature/juvenile animals may provide results that are unrepresentative of mature stock harvested for human consumption.

For each analysis the number of individual shellfish stipulated in the table below should be submitted as a minimum.

SHELLFISH SPECIES AND REQUIRED ANIMAL NUMBERS

Species	Minimum number recommended		
Pacific Oysters Magallana (Crassostrea) gigas	12		
Native Oysters Ostrea edulis	12		
Mussels Mytilus spp.	12		
King Scallops Pecten maximus	12		
Queen Scallops Aequipecten opercularis	12		
Cockles Cerastoderma spp.	24		
Hard Clams Mercenaria mercenaria	12		
Palourdes Ruditapes (Tapes) decussatus	12		
Manila Clams Ruditapes (Tapes) philippinarum	12		
Surf Clams Spisula solida	24		

Important notes:

- The laboratory will only accept and test the species of bivalve shellfish indicated in the table above, Samples consisting of other species of bivalve shellfish will be rejected on arrival, unless exception has been agreed with Cefas in advance of the sample(s) being submitted.
- To enable testing to be carried out in compliance with the international standard method ISO 15216-1:2017, we require a minimum sample size of 10 animals. To allow for removal of animals in poor condition prior to testing we recommend submitting at least 12 animals. In addition, a minimum of 2 grams of digestive tissues is required for testing according to ISO 15216-1:2017 for smaller species a larger number of animals may be required.

Recommended numbers of bivalve shellfish animals to submit for each species are shown in the table above.

If insufficient animals are available to submit, testing may still be possible subject
to prior arrangement with Cefas. For advice, please contact cst@cefas.gov.uk
before shipping material.

Packaging and Transport:

Customers should send samples using an overnight courier service to ensure that samples arrive at the laboratory in a suitable condition for testing. Ideally for live shellfish, samples should be transported in a cool box or similar and kept between 1 $^{\circ}$ C to 10 $^{\circ}$ C during transit. If frozen, samples should be transported in a way that guarantees they do not defrost in transit.

Packaging Guidance

Shellfish to be tested should be placed inside a polythene bag, which is then securely tied (double bagging is recommended to prevent puncture of the bag(s) by the shells). The Sample Submission Form must be fully filled in and placed inside a separate polythene bag and secured to the sample. Both shellfish sample and information sheet should be placed inside the shipping box.

Care must be taken to correctly place any additional cool packs to ensure the shellfish sample does not come into direct contact with the cool pack(s). Should one sample not fit into one box, please split the sample into two boxes, ensuring that each sample is accompanied by a Sample Submission Form (marked 1 of 2 and 2 of 2).

If submitting more than one sample in the same shipping box, each sample must be accompanied by its own Sample Submission Form. Care should be taken to ensure the Sample Submission Form remains attached to the relevant sample so that samples are easily identifiable on arrival at the laboratory.

Once correctly assembled, secure the shipping box lid with adhesive tape to prevent leakage and send the sample via Royal Mail Special Delivery or similar carriers/couriers.

Samples must be delivered to the laboratory as soon as possible and preferably within 24 hours of collection. If short term (up to 24 hours only) storage prior to dispatch/delivery is necessary, samples should be stored between 2 °C and 8 °C. For longer term storage, if necessary, samples can be frozen.

Important notes:

- It is essential that the packaging guidance provided is strictly followed. Failure to adhere to these guidelines may result in the refusal of samples.
- The laboratory will not return transport boxes provided by the customer, unless agreed in advance with the customer. Please note a charge will apply (please consult rate card price list). Cefas will not be held liable for the condition of the transport box once the courier has received it. Any issues relating to damages should be raised with the courier in question.

Submission to the laboratory

Each individual sample must be accompanied with a completed Sample Submission Form. Unlabelled samples arriving at the laboratory with incomplete paperwork or those in poor condition will not be tested. Please mark the samples as "PERISHABLE" Samples should be addressed to:

Cefas Shellfish Testing Molecular Virology Laboratory Barrack Road Weymouth Dorset DT4 8UB UK

It is requested that, where possible, a minimum of one weeks' notice is provided to the laboratory prior to sample submission. If the service's monthly testing capacity has already been reached, customers will be advised not to send samples in for testing. Notice should be in writing to cst@cefas.gov.uk. It is also recommended that you advise the laboratory by emailing cst@cefas.gov.uk when samples are posted as this will help staff identify which samples are due and whether these have been delayed in the post. Please note that samples are only accepted on between the hours of 09:00 and 16:00 on working days.

For samples to be included in the monthly schedule, we ask that samples arrive at the laboratory by the cut off, 12:00 midday, on the identified submission deadline day each month. Samples received after the cut off may be included in the monthly schedule if it is possible for Cefas to do so without impacting turnaround targets for other customers. Please note, this will be at the discretion of Cefas. If this is not possible, customers will be contacted and will be provided with the options to have samples stored according to standard lab procedures and then tested in the schedule for the following month or, to have samples discarded without testing, free of charge.

Important notes:

 Please note that the laboratory is closed on bank holidays and for two weeks during the Christmas period.

Reporting:

Results will be reported electronically to the contact person whose details are provided on the Sample Submission Form. If you require the results to be sent to additional or different contacts, arrangements must be agreed in advance. Please indicate this on the form or contact cst@cefas.gov.uk. Samples must be received at the laboratory by 12:00 pm on the monthly sample submission deadline day to be included in the monthly sampling run. Results for samples included in the monthly sampling run will be reported within 6 working days of the submission deadline subject to satisfactory quality control results.

Submission deadline (12:00)				
April 9 th 2025	Oct 8 th 2025			
May 7 th 2025	Nov 12 th 2025			
Jun 11 th 2025	Dec 10 th 2025			
Jul 9 th 2025	Jan 14 th 2026			
Aug 13 th 2025	Feb 11 th 2026			
Sep 10 th 2025	Mar 11 th 2026			

Reporting deadline (17:00)				
Apr 17 th 2025	Oct 16 th 2025			
May 15 th 2025	Nov 20 th 2025			
Jun 19 th 2025	Dec 18 th 2025			
Jul 17 th 2025	Jan 22 nd 2026			
Aug 21st 2025	Feb 19 th 2026			
Sep 18 th 2025	Mar 19 th 2026			

Sample Retests:

In the event of a retest being required due to quality control issues, reporting within 6 working days of the submission deadline may not be possible. In this instance, the customer will be asked whether they want to proceed with the retest (at no extra cost) or to cancel the test (no fee will be charged). If samples fail quality control on a second attempt, the customer will be charged for the full price of the testing service.

NOTE: samples may fail quality control for several reasons including inherent properties (e.g., presence of high levels of PCR inhibitors).



Sample Submission Form - Virology

Sample reference number *:	
Contact name:	
Sender's address:	
Contact number (landline/mobile):	
Reporting email:	
Date & time of collection:	
Dispatch date:	
Sample quantity:	

* This reference must be unique to each sample and should be a maximum of ten characters. Please ensure you do not refer to the exact geographical origin of your sample.

Analysis required - Tick the box that corresponds to the type of test required & species being tested. Please also indicate whether the sample is shucked or unshucked (S/U) & sample temperature, at time of collection.

sample temperature, at time of collection.				
Species name	Norovirus	Hepatitis A virus	Combined	
Pacific oysters Magallana (Crassostrea) gigas				
Native oysters Ostrea edulis				
Mussels Mytilus spp.				
Cockles Cerastoderma ssp.				
King scallops Pecten maximus				
Queen scallops Aequipecten opercularis				
Hard clams Mercenaria mercenaria				
Surf clams Spisula solida				
Palourde clams Ruditapes (Tapes) decussatus				
Manila clams Ruditapes (Tapes) philippinarum				
Other species - by prior request only				
Please include details of "other species" in box to the right. (include genus & species)				